

基于转录组学分析1-MCP对梨果皮蜡质合成代谢相关基因的影响

于宛婷, 张鑫楠, 孙晓楠, 王文辉, 贾晓辉*

(中国农业科学院果树研究所·农业农村部园艺作物种质资源利用重点实验室·
辽宁省果品贮藏与加工重点实验室, 辽宁兴城 125100)

摘要:【目的】通过分析1-MCP对常温货架期梨果皮蜡质合成代谢基因的影响,为探究1-MCP调控梨果皮油腻化的分子机制提供理论依据。【方法】以玉露香梨(*Pyrus sinkiangensis* 'Yuluxiang')为试材,采用 $1.0 \mu\text{L}\cdot\text{L}^{-1}$ 的1-MCP熏蒸24 h,以未经处理的果实为对照,于20 °C货架21 d,持续观察果实外观变化,每隔7 d取一次梨果皮冻样并用于转录组测序分析,利用RT-qPCR技术验证显著差异基因的表达情况。【结果】货架前14 d,1-MCP处理的果实外观品质明显优于对照,同时果面亮度L值较对照更低;转录组测序结果表明,相比于货架0 d和14 d,两组梨果在货架第7天上调和下调表达的显著差异基因数均最多,且此时期果实外观差异最明显;富集分析结果显示,在货架第7天,共95个显著差异基因富集到脂质代谢通路,这些基因参与脂肪酸的延伸、合成和降解等脂质次级代谢途径;RT-qPCR验证结果表明,与对照相比,1-MCP处理显著抑制货架7 d、14 d的*PyLACS9*、*PyKCS20*和*PyCER1*基因的上调表达,抑制货架第7天的*PyPLDALPHA4*和*PyFAD2*基因的上调表达,通过上述验证了1-MCP对玉露香梨果皮蜡质合成代谢基因表达模式的调控作用;*PyLACS9*与果面亮度L值呈显著正相关($r=0.99, p<0.05$),因此推测*PyLACS9*可能是导致常温贮藏下玉露香梨果皮油腻化的关键基因。【结论】1-MCP处理能够维持常温货架期玉露香梨果实较好的外观品质和较低的L值,1-MCP可能通过调控果皮蜡质合成代谢基因的表达水平进而抑制果皮油腻化。

关键词:玉露香梨;油腻化;1-甲基环丙烯;转录组;蜡质合成代谢基因

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Effects of 1-MCP on the genes related to wax anabolism in pear peel based on transcriptomics

YU Wanting, ZHANG Xinnan, SUN Xiaonan, WANG Wenhui, JIA Xiaohui*

(Research Institute of Pomology, Chinese Academy of Agricultural Sciences/Key Laboratory of Germplasm Resources Utilization of Horticultural Crops, Ministry of Agriculture and Rural Affairs/Key Laboratory of Fruits Storage and Processing, Xingcheng 125100, Liaoning, China)

Abstract: 【Objective】According to the investigation and laboratory research of our study group for several years, it has been found that the main pear cultivar Yuluxiang was easy to become greasy in the early stage of shelf life at room temperature, and also in the middle and late stage of cold storage. Previous researchers have reported that the expression levels of genes related to wax anabolism in the peel and ethylene synthesis are related closely to the degree of oiliness in the peel. Moreover, the inhibitive effect of 1-MCP on ethylene release has been widely reported, and thus the regulation mode of 1-MCP on the wax synthesis and metabolism genes in pear peel during shelf life at room temperature was studied, which provided a theoretical basis for a preliminary exploration of the molecular mechanism of 1-MCP regulation of pear peel greasiness. 【Methods】Yuluxiang pear was used as the experimental material,

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作者简介:于宛婷,女,在读博士研究生,主要从事果品贮藏保鲜与采后生理研究。E-mail:abcdefg1468929@qq.com

*通信作者 Author for correspondence. E-mail:Jiaxiaohui@caas.cn

and fruits with uniform size and without pests, diseases and bumps were selected as experimental samples, which were fumigated with $1.0 \mu\text{L} \cdot \text{L}^{-1}$ 1-MCP for 24 h, and the untreated fruit was used as the control. Then, the appearance changes of the fruit were observed on the shelf at $20 \text{ }^{\circ}\text{C}$ for 14 days. At the same time, the fruit surface brightness L value was detected. RNA was extracted from frozen peel samples every 7 days for transcriptome sequencing and data analysis, and RT-qPCR technology was used to verify the significant difference genes. **【Results】** During the whole shelf life, the appearance quality of fruit treated with 1-MCP was better than that of the control. On the 14th day of shelf life, it could still maintain a better green fruit surface, and a lower L value of fruit surface brightness, and there was no greasy phenomenon on the fruit surface at this time. Through transcriptome sequencing and data analysis, a total of 103283229900 clean reads were obtained, and the data quality was high. The results of reference genome alignment showed that the sequencing data of the Yuluxiang fruit peel were well aligned with the pear reference genome. Compared with the control, there were 2463 differentially expressed genes up-regulated and 599 differentially expressed genes down-regulated in the 1-MCP group on the 7th day of shelf life. On the 14th day of shelf life, there were 786 differentially expressed genes up-regulated and 284 differentially expressed genes down-regulated in the 1-MCP group. The number of differentially expressed genes in two groups was the highest on the 7th day of shelf life, indicating that the difference between the two groups was large. Therefore, the significant differential genes of two groups of fruits on the 7th day of shelf life were enriched into the lipid metabolism pathway through KEGG, a total of 95 significant differential genes, and 13 secondary lipid metabolism pathways were enriched into this pathway. The secondary metabolic pathways directly related to wax biosynthesis were biosynthesis of cutin, suberine, wax, fatty acid elongation, fatty acid biosynthesis and fatty acid degradation. In this study, a total of 7 significantly different genes (*PyKCSI*, *PyKCS20*, *PyKCR1*, *PyPLDALPHA4*, *PyLACS9*, *PyFAD2* and *PyCER1*) were enriched in lipid metabolism pathways, which were involved in lipid secondary metabolic pathways such as fatty acid extension, synthesis and degradation, and *PyKCS20*, *PyKCR1*, *PyPLDALPHA4*, *PyLACS9*, *PyFAD2* and *PyCER1* were down-regulated and *PyKCSI* was up-regulated in the 1-MCP-treated group. The results of RT-qPCR showed that 1-MCP treatment significantly inhibited the up-regulated expression of *PyLACS9*, *PyKCS20* and *PyCER1* genes during the whole shelf life, and inhibited the up-regulated expression of *PyPLDALPHA4* and *PyFAD2* genes in the first 7 days of shelf life, which verified the regulation of 1-MCP on the expression pattern of wax anabolism genes in the peel of Yuluxiang pear. The results of correlation analysis showed that *PyLACS9* was significantly and positively correlated with the L value of fruit surface brightness ($p < 0.05$), and the L value was strongly positively correlated with *PyKCS20*, strongly negatively correlated with *PyPLDALPHA4* and *PyACS-1*, negatively correlated with *PyKCSI*, and weakly correlated with *PyKCR1*, *PyCER1* and *PyACO2*, but not significantly ($p \geq 0.05$). In addition, the correlation between different differential genes showed that *LACS9* was strongly negatively correlated with *PyACS-1*, *PyKCR1* and *PyCER1* were strongly positively correlated with *PyACO2*, *PyPLDALPHA4* was strongly positively correlated with *PyACS-1*, *PyFAD2* was strongly positively correlated with *PyACO2* and *PyACS-1*, and they were not significant ($p \geq 0.05$), so it was speculated that *PyLACS9* may be the key gene leading to the greasiness of Yuluxiang fruit peel. **【Conclusion】** In summary, 1-MCP treatment could maintain the good appearance quality and low L value of Yuluxiang fruit during shelf life at room temperature, which may affect the greasiness of peel by affecting the expression level of wax anabolism genes in Yuluxiang fruit peel. Exploring the regulatory effect of 1-MCP on the waxy synthesis and metabolism genes in the peel of Yuluxiang pear during shelf life provides a theoretical basis for the prevention and control of post-

harvest greasiness of pear fruit.

Key words: Yuluxiang pear; Greasiness; 1-methylcyclopropene; Transcriptome; Waxy anabolic genes

水果表皮覆盖的蜡质层能够抑制果实表面渗透性、减少蒸腾作用以及防止微生物入侵^[1-2]。表皮蜡质的合成底物为具有16或18个碳原子的脂肪酸,其主要在细胞质(C16)或质体(C16、C18)中合成。脂肪酸在长链脂酰辅酶A(long-chain-acyl-coenzyme A, LACS)的作用下分别转化为16和18个碳原子的脂酰基辅酶A。在内质网中,经脂肪酸伸长酶复合物(β -酮脂酰辅酶A合酶, β -ketoacyl-acyl-coenzyme A synthase, KCS; β -酮脂酰辅酶A还原酶, β -ketoacyl-acyl-coenzyme A reductase, KCR; β -羟脂酰辅酶A脱水酶, β -hydroxy-acyl-coenzyme A dehydrase, HCD;烯脂酰辅酶A脱水酶, enoyl-CoA reductase, ECR)催化,延伸成非常长链脂肪酸(Very long chain fatty acids, VLCFAs)。VLCFAs又通过酰基还原途径形成伯醇,并通过脱羧基化生成其他组分,如醛、烷烃、仲醇和酮等,最后这些蜡组分穿过细胞壁,到达果实角质层,并进行自我组装形成肉眼可见的白色霜状或油渍状物质^[3-6]。

水果在贮藏过程中,果皮蜡质组分由于受到基因的调控而发生性质的改变。Wu等^[7]研究了3个亚洲梨品种(库尔勒香梨、雪花梨和玉露香梨)在贮藏过程中表皮蜡质合成代谢关键基因的表达模式,结果表明,果实中*PyCER6*、*PyKCS9*、*PyKCS20*和*PyFDH1*基因上调表达,*PyCER60*、*PyDGAT1*和*PyMAH1*等基因下调表达,这些基因均参与了果皮蜡质的合成。贮藏期果皮蜡质性质的变化还可能导致果实外观品质下降,即果皮油腻化现象。目前果皮油腻化被认为是一种果实采后常见的生理病害,在苹果和梨上均有发生^[8-12],这种病害会影响果实外观品质进而降低商品价值^[13]。根据笔者课题组多年生产实践调研和实验室研究发现,梨主栽品种玉露香(*Pyrus sinkiangensis* 'Yuluxiang')在常温货架前期和冷藏中后期较易油腻化^[12,14-16]。已有研究报道,果皮蜡质合成代谢和乙烯合成相关基因的表达水平、乙烯释放量与果皮油腻化程度密切相关^[17,18]。Jiang等^[19]研究表明,苹果的*MdFAD27*和*MdFAD28*基因参与了果实酯类底物和油腻化外观的形成,且果实的油腻化程度与其乙烯释放速率显著相关。因此,研究梨果皮蜡质合成代谢途径及乙烯合成基因的表达情况对探

究梨果皮油腻化发生的分子机制具有重要意义。

1-MCP是一种乙烯抑制剂,它通过与乙烯竞争受体结合位点而抑制乙烯的释放。1-MCP已广泛应用于果蔬采后贮藏保鲜。Yang等^[20]研究表明,库尔勒香梨的果实经1-MCP处理后,与长链蜡质合成相关基因的上调表达被抑制。笔者实验室前期研究发现,1-MCP处理可以抑制常温货架期玉露香梨果皮多种蜡质组分包含醇类、醛类、脂肪酸类和烯炔类化合物含量的上升^[21]。因此,挖掘1-MCP调控玉露香梨果皮蜡质合成的关键基因,对阐明1-MCP调控果皮油腻化潜在的分子机制具有重要意义。本研究中,采用转录组测序技术,以玉露香梨为试材,采用 $1.0 \mu\text{L} \cdot \text{L}^{-1}$ 1-MCP熏蒸处理24 h,在20 °C条件下贮藏0、7、14 d,筛选不同时间处理后果皮的差异表达基因,旨在探究1-MCP对货架期玉露香梨果皮蜡质合成代谢基因的调控作用,为梨果采后油腻化防控提供理论依据。

1 材料和方法

1.1 试验材料与处理

供试玉露香梨采摘于辽宁省葫芦岛市赵家沟村,果园管理中上等水平,树龄10 a(年),土壤为壤砂土,采收当年盛花期为4月15日,采收时间为商业采收期9月15日,采摘后于2 h内运送至中国农业科学院果树研究所。挑选大小均一、无病虫害和磕碰伤的果实用于后续试验。采用 $1.0 \mu\text{L} \cdot \text{L}^{-1}$ 的1-MCP在室温(20±1)°C条件下熏蒸24 h,以未熏蒸果实为对照,于20 °C环境下放置21 d,每隔7 d取1次玉露香梨果皮,于-80 °C超低温冰箱冻存,用于后续分析。

1.2 果面亮度L值

果面亮度L值的检测参照于宛婷等^[22]的方法。

1.3 RNA提取与cDNA合成

采用RN53-EASYspin Plus多糖多酚复杂植物RNA快速提取试剂盒(艾德莱,北京贝洛生物科技有限公司)提取试验梨果皮的总RNA。利用1%琼脂糖凝胶电泳和超微量分光光度计检测RNA的完整性、纯度和浓度。将检测合格后(RNA无显著降解, A_{260}/A_{280} 在2.0~2.2之间)的样品送至广州基迪奥生物科技有限公司进行文库构建和转录组测序工

作;另使用 cDNA 合成试剂盒(Thermo Scientific™ EP0733,北京中泰弘丰科技有限公司)将 RNA 反转录成 cDNA,用于后续研究。

1.4 转录组数据质控和参考基因组比对分析

所选样品的时间为货架 0、7、14 d,每个处理设置 3 个生物学重复。通过对玉露香梨果皮测序得到的原始数据进行数据过滤,以减少无效数据造成的分析干扰。首先对下机的 raw reads 利用 fastp^[23]进行质控,具体包括去除含接头(adapter)的 reads、含 N(N 表示无法确定的碱基信息)比例大于 10%的 reads、含 100% A 碱基的 reads、低质量 reads(质量值 $Q \leq 20$ 的碱基数占整条 read 的 50%以上),最终得到 clean reads。并通过 GC 含量、Q20、Q30 数据指标对 clean reads 进行评判,得到的 clean reads 用于后续转录组分析。玉露香梨的参考基因组和基因模型注释文件从 NCBI 网站(https://www.ncbi.nlm.nih.gov/genomes/all/GCF_000315295.1)下载。采用 HISTA2^[24]软件开展基于参考基因组的比对分析,通过全局和局部搜索比对到 RNA-Seq 测序数据中的 spliced reads。

1.5 基因的定量分析和差异表达基因的筛选

基因表达量的准确性依赖于转录本重构结果的

完善程度。根据 HISTA2 的比对结果,利用 Stringtie^[25]重构转录本,并利用 RSEM^[26]计算每个样本中所有基因的表达量。使用 FPKM(Fragments Per Kilobase of exon model per Million mapped fragments)矫正测序深度和基因长度对表达量的影响。在基因表达量的基础上通过 DESeq2 进行差异表达分析,差异表达基因的筛选标准为基因表达量变化倍数 $|\log_2 \text{fold change}| > 1$ 和 FDR 值(false discovery rate) < 0.05 。

1.6 差异表达基因的富集分析

应用 Omicsmart-组学挖掘数据平台(<https://www.omicsmart.com/>)对差异表达基因进行 KEGG(Kyoto Encyclopedia of Genes and Genomes)通路富集分析。KEGG 通路富集以 corrected-*p* value ≤ 0.05 作为显著性富集的阈值,将差异表达基因筛选、分类为不同的代谢通路和次级代谢途径。

1.7 RT-qPCR 验证分析

根据 1.6 脂质代谢途径共筛选出 7 个蜡质合成相关显著差异基因,另筛选 2 个乙烯合成显著差异基因,设计引物(表 1),根据文献报道的亚洲梨内参基因^[7]设计内参引物进行 RT-qPCR 验证。引物均采购自生工生物工程(沈阳)股份有限公司。利用荧光

表 1 RT-qPCR 实验所用引物
Table 1 Primers used in RT-qPCR

基因名称 Gene name	基因序列号 Gene bank accession	引物编码 Primer code	引物序列(5'-3') Primer sequence (5'-3')
Reference	Pbr035952.1	Reference-F Reference-R	GCAGTTTCGTGAGTGTAGAGGA ATCTGGGCGTTCTCGTTTCA
<i>PyLACS9</i>	LOC103953155	LACS9-F LACS9-R	AAACTTCAGCACGGGGAGTA AGTGATTCTTGTTCAGCC
<i>PyKCS20</i>	LOC103929791	KCS20-F KCS20-R	GCAGAATCAGAACCAGGGAGA CTGTGAGCGTAGAGAGGTGG
<i>PyACO2</i>	LOC103946813	ACO2-F ACO2-R	CCCAATGCACCACTCCAT CTTGGGATAAGTTGGGGCGT
<i>PyACS-1</i>	LOC103931583	ACS-1-F ACS-1-R	GGCTTTTAGCTCCCCATCCT CGTCGTTGGAGTAAATGGCG
<i>PyKCS1</i>	LOC103956371	KCS1-F KCS1-R	AGCGAGAAGATGACAAGGGG GTACGGTTTGACCTTCGCCT
<i>PyPLDALPH4</i>	LOC103957160	PLDALPH4-F PLDALPH4-R	TGGACGGACATCGAGACACT TATGCTCCAATGTTGCCTCCC
<i>PyFAD2</i>	LOC103958977	FAD2-F FAD2-R	ATCCTCCACTCTTGCTCCT GGCCAGCCTAGAGTGAGTTG
<i>PyCER1</i>	LOC103965506	CER1-F CER1-R	CTCGCTACCACTCTCATCACC CAGCATAGACTGCGACAGAAG
<i>PyKCR1</i>	LOC103949861	KCR1-F KCR1-R	TAAAAGGGCTTGACGTGGGC CTCAGCATAACCGGAAGCACT

定量 PCR 仪 (BioRAD CFX96 Touch, USA), 采用 Taq Pro Universal SYBR qPCR Master Mix 试剂盒的方法测定上述目标基因的相对丰度。qPCR 反应条件如下: 95 °C 保持 30 s 以激活 Taq 酶, 随后 95 °C 变性 5 s, 54 °C 退火及延伸 30 s 并循环 40 次。溶解曲线采集程序为: 95 °C, 15 s; 60 °C, 60 s; 95 °C, 15 s。最后, 根据扩增反应得到相应的 Ct 值, 采用 $-2^{-\Delta\Delta Ct}$ 的方法计算目标基因的相对表达量。

1.8 数据分析

利用 Microsoft Excel 2016 软件绘图、计算平均值和标准误差。利用 SPSS 25.0 软件进行方差分析, 采用邓肯法检验组间的差异性。

2 结果与分析

2.1 1-MCP 对玉露香梨货架期果皮颜色和油腻化的影响

如图 1 所示, 随着货架时间的延长, 玉露香梨果面绿色逐渐褪去, 逐渐转黄且越来越亮, 同时伴随油腻化的发生, 而 1-MCP 处理显著抑制了货架前 7 d 果皮油腻化的发生。货架第 14 天, 对照组梨果面基本全部转黄且油腻化现象也更加严重, 而 1-MCP 处理组仍保持较好的绿色。货架第 21 天, 对照和 1-MCP 组梨果面均全部转黄, 用手摩擦有油腻感, 已失去商品价值。

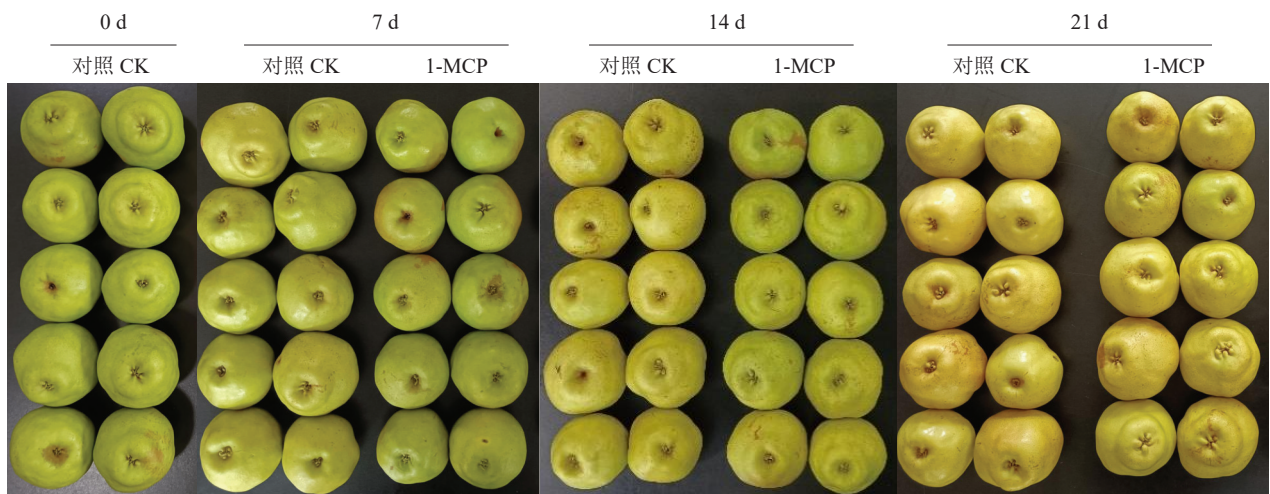


图 1 1-MCP 处理对玉露香梨货架期果皮颜色和油腻化的影响

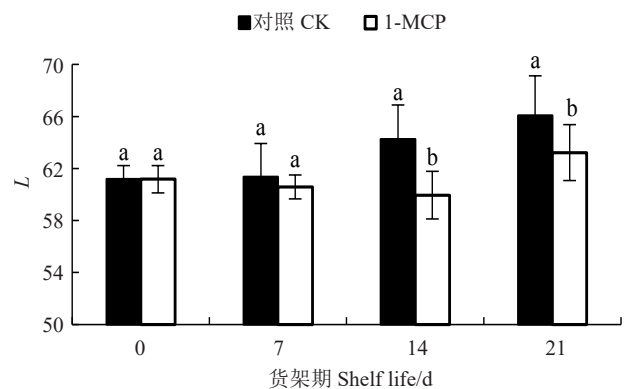
Fig. 1 Effects of 1-MCP treatment on Yuluxiang pears' appearance and greasy peel during shelf life

2.2 1-MCP 对玉露香梨货架期果面 L 值的影响

L 值可反映果皮油腻化的程度, L 值越高, 果皮越油腻^[21, 27]。如图 2 所示, 随着货架时间的延长, 对照组 L 值不断上升, 而 1-MCP 处理组 L 值变化不明显, 且在货架第 14 天较前期下降。货架第 7 天, 对照组果实 L 值高于 1-MCP 处理组, 但两组差异不显著 ($p \geq 0.05$)。货架第 14 天和第 21 天, 对照组果实 L 值均显著高于 1-MCP 处理 ($p < 0.05$)。

2.3 转录组测序数据质量评价

分别对货架 0、7、14 d 的玉露香梨果皮进行转录组测序分析, 5 组处理、每组 3 次重复, 构建了 15 个 cDNA 文库, 共获得 103 283 229 900 条 clean reads, 其中 Q20 所占百分比均在 97.90% 以上, Q30 所占百分比均在 93.80% 以上, 平均每个样品的 GC 含量为 46.03% (表 2)。由此可知, 本次测序得到的



不同小写字母表示在 0.05 水平上差异显著。

Different small letters indicate significant differences at the 0.05 level.

图 2 1-MCP 处理对玉露香梨货架期果面 L 值的影响

Fig. 2 Effect of 1-MCP treatment on L value of Yuluxiang pear during shelf life

表 2 玉露香梨果皮转录组测序数据质量信息

Table 2 Quality information of transcriptome sequencing data of Yuluxiang pear

样品编号 Sample number	总原始序列 Total raw reads	质控后序列总数 Total clean reads	Q20/%	Q30/%	GC/%
0d-1	6 119 576 100	6 049 361 071	98.26	94.65	46.00
0d-2	5 969 966 400	5 896 628 694	98.40	95.04	46.18
0d-3	5 940 231 600	5 874 575 612	98.45	95.16	46.01
CK1-7d	6 469 605 900	6 388 696 304	98.40	95.02	46.19
CK2-7d	6 800 348 400	6 735 529 713	98.07	94.18	46.08
CK3-7d	6 346 413 900	6 283 227 060	98.11	94.29	46.05
TR1-7d	7 839 364 200	7 758 500 798	98.67	95.81	46.04
TR2-7d	8 217 298 800	8 122 916 028	98.45	95.18	45.94
TR3-7d	8 611 986 900	8 519 660 224	98.16	94.40	45.88
CK1-14d	6 500 761 500	6 427 653 280	98.49	95.33	45.94
CK2-14d	6 701 087 400	6 617 327 089	98.13	94.32	46.03
CK3-14d	6 839 873 700	6 758 208 600	98.38	95.04	46.10
TR1-14d	6 937 250 400	6 849 549 316	98.05	94.17	45.99
TR2-14d	7 194 605 400	7 102 762 548	98.64	95.73	45.97
TR3-14d	6 794 859 300	6 724 750 573	97.95	93.88	46.00

注:TR 代表 1-MCP 处理,下同。Q20、Q30 表示 Phred 数值大于 20、30 的碱基占总碱基数的百分比;GC 表示碱基 G 和 C 的数量总和占总碱基数量的百分比。

Note: TR represents 1-MCP processing, the same below. Q20 and Q30 represent the percentage of bases with Phred values greater than 20 and 30 in the total number of bases. GC represents the total number of bases G and C as a percentage of the total number of bases.

玉露香梨果皮转录组的数据量和质量都较高,可满足下一步基因组比对分析的要求。

将检验合格的 clean reads 与梨参考基因组进行比对分析,结果(表3)显示,能定位到参考基因组上的测序序列的占比(mapping rate)范围在 76.45%~78.61%,均大于 70%,表明选择的梨基因组合适,且梨样品不存在污染。在参考序列上有唯一比对(uniqely mapping)位置的 clean reads 的占比范围在 68.56%~70.79%。在参考序列上有多个比对位置(multiple mapping)的 clean reads 的占比范围在 7.73%~8.09%,均小于 10%。上述结果表明,玉露香梨果皮的转录组测序数据与梨参考基因组比对结果良好,可用于下一步差异表达基因的分析。

2.4 货架期玉露香梨差异表达基因分析

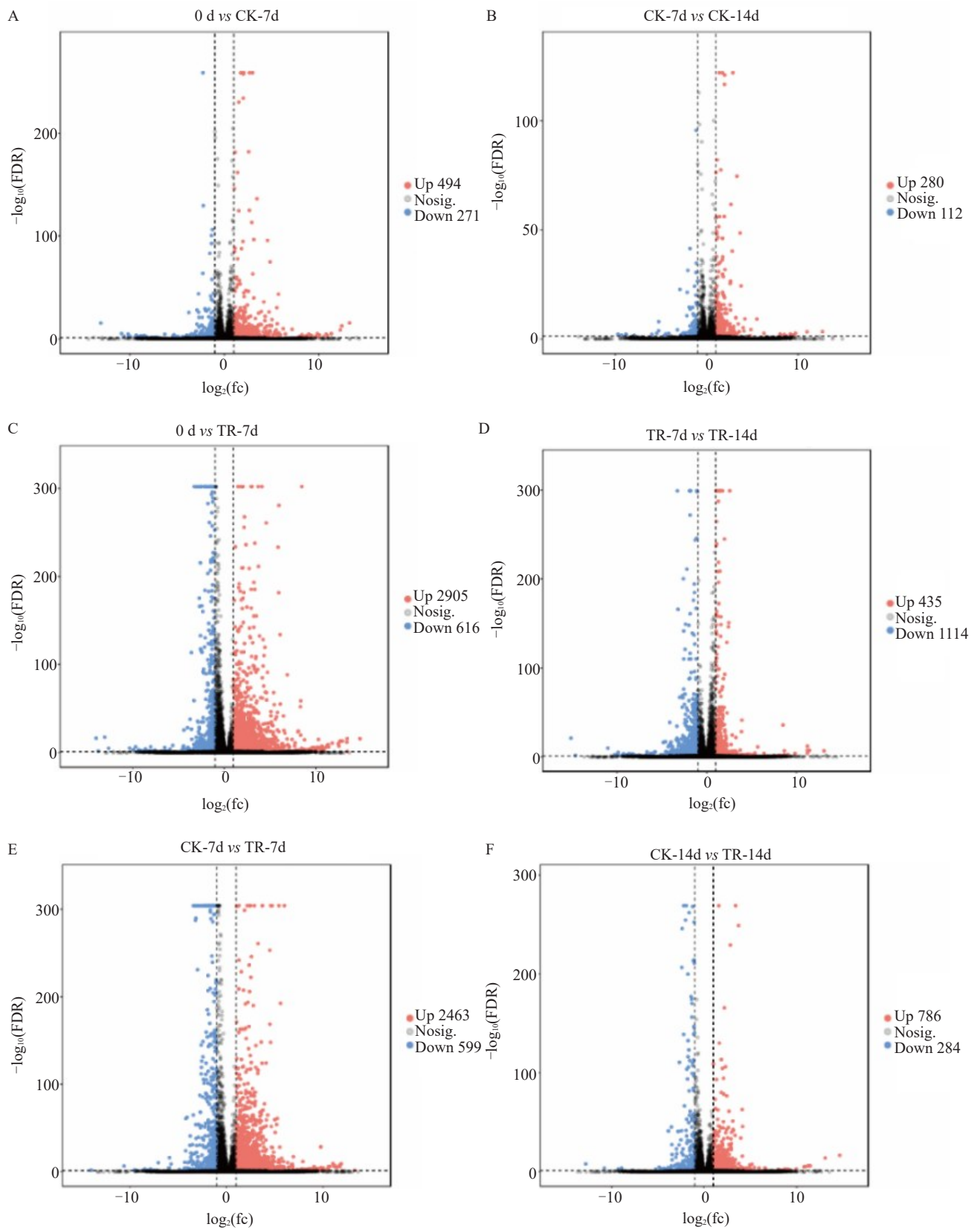
由图 3 可知,与货架 0 d 相比,货架 7 d 后,对照组共有 494 个差异基因上调表达,271 个差异基因下调表达,1-MCP 组共有 2905 个差异基因上调表达,616 个基因下调表达(图 3-A、C)。与货架 7 d 相比,货架 14 d 后,对照组共有 280 个差异基因上调表达,112 个差异基因下调表达,1-MCP 组共有 435 个差异基因上调表达,1114 个差异基因下调表达(图 3-B、D)。此外,对比同一货架期不同处理的差异基因表

表 3 玉露香梨参考基因组比对情况统计

Table 3 Reference genome comparison statistics of Yuluxiang pear

样品名称 Sample name	总比对率 Total mapping rate/%	唯一比对率 Uniquely mapping rate/%	多重比对率 Multiple mapping rate/%
0d-1	78.31	70.55	7.76
0d-2	78.37	70.49	7.88
0d-3	78.52	70.79	7.73
CK1-7d	78.46	70.42	8.04
CK2-7d	78.29	70.38	7.91
CK3-7d	78.29	70.38	7.92
TR1-7d	77.44	69.51	7.93
TR2-7d	77.53	69.65	7.88
TR3-7d	77.16	69.31	7.85
CK1-14d	76.45	68.56	7.89
CK2-14d	76.58	68.70	7.88
CK3-14d	76.74	68.76	7.98
TR1-14d	78.27	70.31	7.96
TR2-14d	78.61	70.52	8.09
TR3-14d	78.17	70.33	7.84

达情况发现,与对照组相比,货架 7 d 后,1-MCP 组共有 2463 个差异基因上调表达,599 个差异基因下调表达(图 3-E),货架 14 d 后,1-MCP 组共有 786 个



红色代表显著上调表达基因,蓝色表示显著下调表达基因。

Red represents the significantly up-regulated genes, and blue represents the significantly down-regulated genes.

图3 常温货架期 1-MCP 处理的玉露香梨差异表达基因火山图

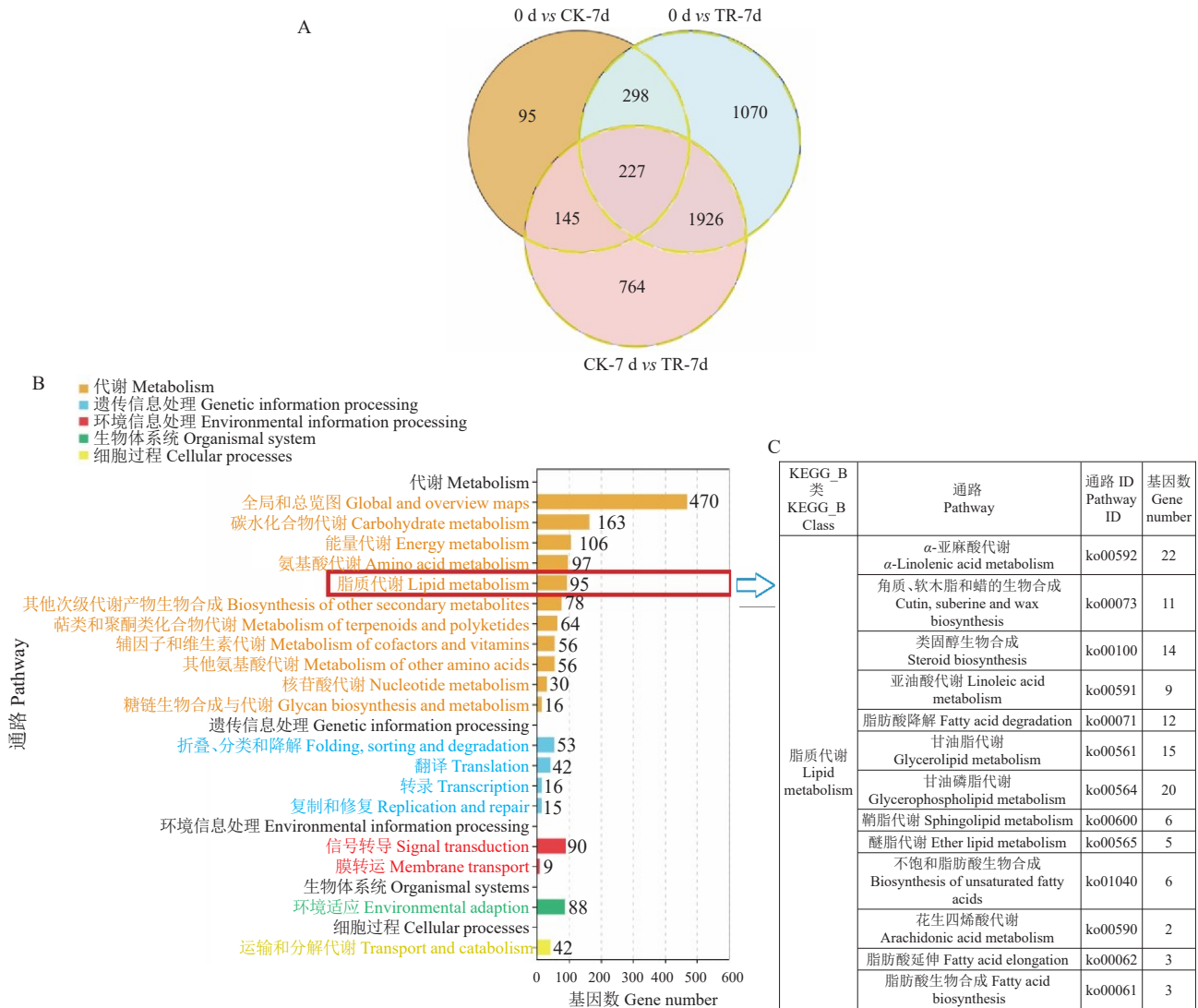
Fig. 3 Volcano map of differentially expressed genes of Yuluxiang pear treated with 1-MCP at room temperature during shelf life

差异基因上调表达,284个差异基因下调表达(图3-F)。因为货架7 d后,1-MCP处理组玉露香梨果实显著上调表达的差异基因数远高于对照组,所以在后续分析中重点关注此时间点。

2.5 KEGG 富集分析玉露香梨脂质代谢通路和次级代谢途径

对货架前7 d的对照和处理组玉露香梨果皮显著差异基因进行韦恩图分析,如图4-A所示,而后将除0 d vs CK-7d的95个基因之外的所有显著差异基因进行KEGG富集分析,结果如图4-B所示,货架第

7天,共95个差异基因富集到脂质代谢通路。这些差异基因主要富集在角质(cutin)、软木脂(suberine)和蜡(wax)的生物合成,倍半萜和三萜类生物合成, α -亚麻酸和亚油酸代谢,甘油酯代谢,脂肪酸延伸(fatty acid elongation),脂肪酸生物合成和降解(fatty acid biosynthesis and degradation),不饱和脂肪酸的生物合成等13个次级脂质代谢途径。根据已有文献报道^[28]与蜡质合成代谢直接相关的次级代谢途径分别为角质、软木脂和蜡的生物合成、脂肪酸延伸、脂肪酸生物合成和脂肪酸降解。在本研究中,玉露香



A. 货架0 d、7 d对照和处理组显著差异基因韦恩图; B. 对照和处理组在货架7 d以及1-MCP在货架0 d和7 d的显著差异基因集的KEGG富集分析图; C. 对照和处理组在不同脂质代谢途径下的显著差异基因数。

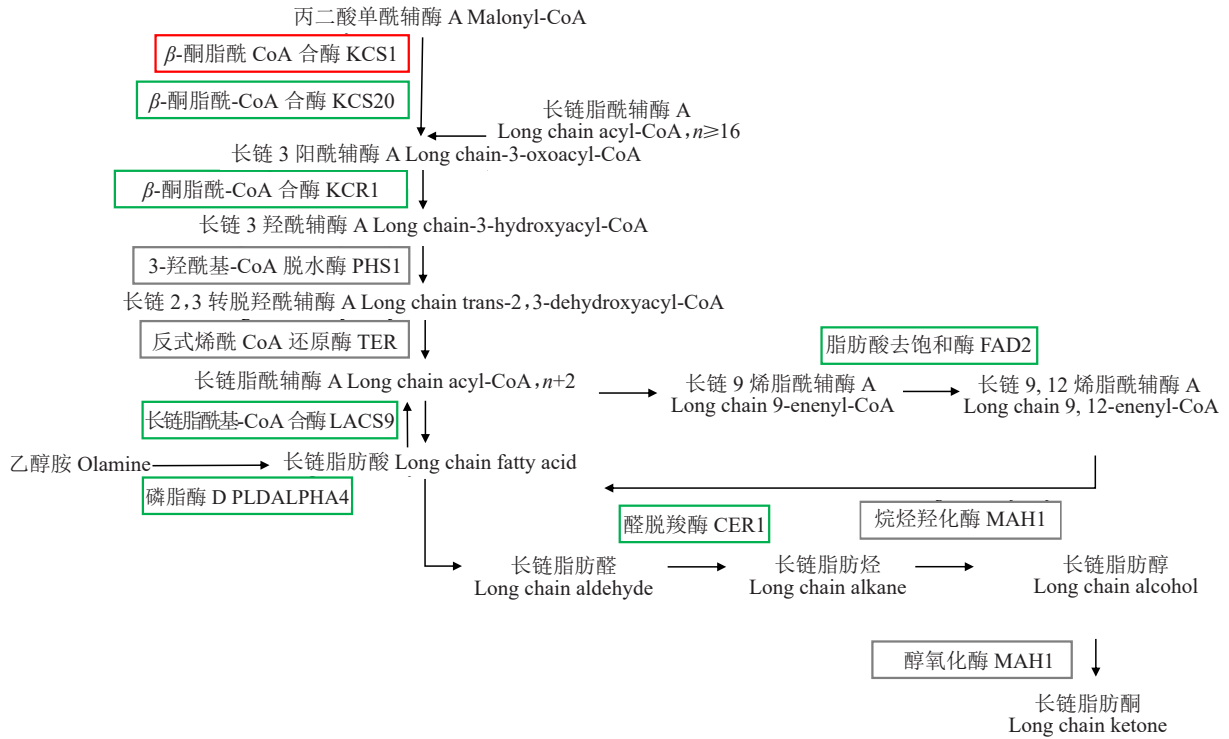
A. The significant difference gene Venn diagram of the control and treatment group on the 0 d and 7 d of the shelf; B. The KEGG enrichment analysis diagram of the significant difference gene set of the control and treatment group on the 7th day of the shelf and 1-MCP on the 0th and 7th day of the shelf; C. The number of significantly different genes in the control and treatment group under different lipid metabolism pathways.

图 4 差异基因韦恩图分析和 KEGG 富集分析次级脂质代谢途径 (0~7 d)

Fig. 4 Venn diagram analysis of differential expressed gene and KEGG enrichment analysis of secondary lipid metabolic pathway (0~7 d)

梨果皮转录组测序分析结果显示,在与蜡质合成直接相关的途径中共检测到29个差异表达转录本,包括角质、软木脂和蜡的生物合成、脂肪酸降解、脂肪酸生物合成和脂肪酸延伸途径的11、12、3、3个差异表达基因。

对货架7 d后对照组和1-MCP处理的玉露香梨果皮蜡质合成脂质代谢通路图进行分析(图5),共检测到7个参与脂肪酸延伸途径、脂肪酸脱羧途径和酰基还原途径的差异表达转录本,分别为*PyKCSI*、*PyKCS20*、*PyKCR1*、*PyPLDALPHA4*、*PyLACS9*、*Py-*



红色方框表示上调表达基因,绿色方框表示下调表达基因。

The red box indicates the up-regulated genes and the green box indicates the down-regulated genes.

图5 玉露香梨果脂质代谢通路图

Fig. 5 Lipid metabolism pathway map of Yuluxiang pear peel

FAD2、*PyCER1*。相比于对照,1-MCP处理导致*PyKCSI*上调表达,*PyKCS20*、*PyKCR1*、*PyCER1*、*PyLACS9*、*PyPLDALPHA4*和*PyFAD2*下调表达。

2.6 RT-qPCR验证

根据KEGG富集分析结果,选择7个与脂质合成代谢相关的基因和2个乙烯合成相关基因进行RT-qPCR验证。结果表明,与对照相比,货架7 d和14 d后,1-MCP处理均显著抑制*PyLACS9*、*PyKCS20*、*PyCER1*、*PyACO2*和*PyACS-1*基因的表达,显著促进*PyKCSI*基因的表达;货架7 d后,1-MCP处理显著抑制*PyPLDALPHA4*和*PyFAD2*基因的表达(图6)。

2.7 差异基因相对表达量与L值的相关性分析

如表4所示,L值与*PyLACS9*呈显著正相关($r=0.99, p<0.05$),与*PyKCS20*呈正相关,与*PyKCSI*、

PyPLDALPHA4、*PyACS-1*呈负相关,与*PyKCR1*、*PyCER1*、*PyACO2*呈弱相关。此外,不同差异基因间相关性表现为,*PyLACS9*与*PyACS-1*呈负相关,*PyKCR1*、*PyCER1*与*PyACO2*呈正相关,*PyPLDALPHA4*与*PyACS-1*呈正相关,*PyFAD2*与*PyACO2*、*PyACS-1*均呈负相关。

3 讨论

3.1 果皮蜡质合成代谢与油腻化的关系

蜡质组分的合成代谢反应在采前植物的果实、叶片和茎以及采后贮藏的果实中均会发生^[29]。植物在生长过程中积累的表皮蜡质具有维持植物体内水分平衡、防止水分流失、保护植物免受病原体入侵和昆虫食草动物的侵袭^[30]的功能,而在果实贮藏期间,果皮蜡质合成代谢途径的变化可能会导致果实外观

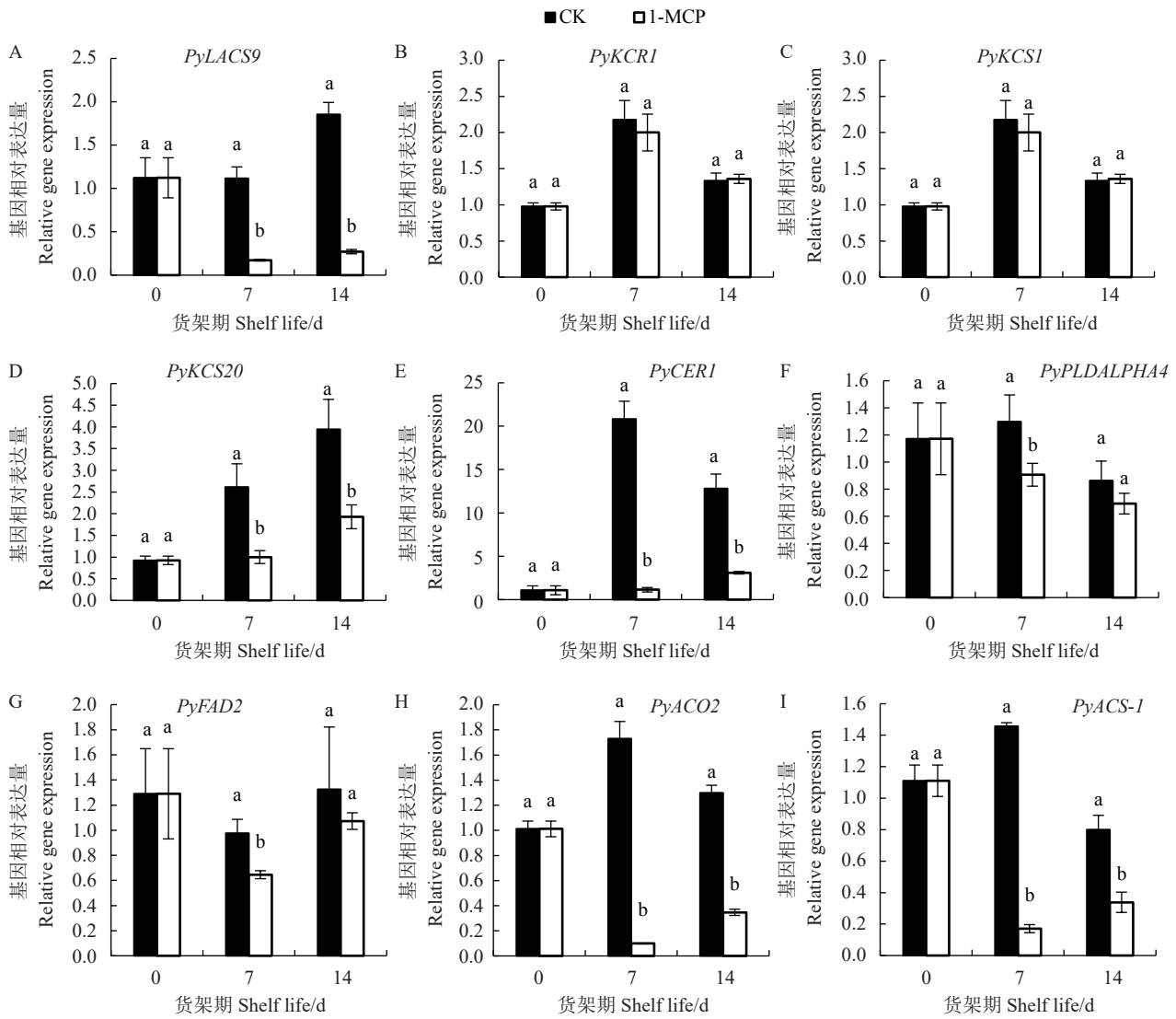


图 6 RT-qPCR 验证 1-MCP 处理的玉露香梨显著差异表达基因

Fig. 6 RT-qPCR verification of significant differentially expressed genes of 1-MCP treated Yuluxiang pear

表 4 1-MCP 处理的玉露香梨显著差异基因与果面亮度 L 值的相关性分析

Table 4 Correlation analysis of significantly differential expressed genes and fruit surface brightness L value of 1-MCP treated Yuluxiang pear

基因或 L 值 Genes or L value	<i>PyLACS9</i>	<i>PyKCR1</i>	<i>PyKCS1</i>	<i>PyKCS20</i>	<i>PyCER1</i>	<i>PyPLDALPHA4</i>	<i>PyFAD2</i>	<i>PyACO2</i>	<i>PyACS-1</i>	L 值 L value
<i>PyLACS9</i>	1.00									
<i>PyKCR1</i>	-0.23	1.00								
<i>PyKCS1</i>	-0.72	-0.51	1.00							
<i>PyKCS20</i>	0.83	0.35	-0.98	1.00						
<i>PyCER1</i>	0.10	0.94	-0.77	0.64	1.00					
<i>PyPLDALPHA4</i>	-0.96	0.49	0.49	-0.64	0.18	1.00				
<i>PyFAD2</i>	0.58	-0.93	0.15	0.02	-0.75	-0.78	1.00			
<i>PyACO2</i>	-0.12	0.99	-0.61	0.46	0.98	0.39	-0.88	1.00		
<i>PyACS-1</i>	-0.85	0.71	0.25	-0.41	0.43	0.96	-0.92	0.62	1.00	
L 值 L value	0.99*	-0.17	-0.76	0.86	0.17	-0.94	0.53	-0.06	-0.82	1.00

注: *表示在 0.05 水平上相关性显著。

Note: * indicates a significant correlation at the 0.05 level.

变差,如果面的油渍感和黏腻感^[18]。

Li等^[11]以玉露香梨、雪花梨和鸭梨3个亚洲梨品种为试验材料,果实冷藏45 d后,15个表皮蜡质合成代谢和蜡质转运相关基因(*PyLACS1*、*PyKCS2*、*PyKCS6*、*PyFDH*、*PyKCS20*、*PyGL8*、*PyCER10*、*PyCER60*、*PyLTPG1*、*PyLTP4*、*PyABCG12*、*PyCER1L*、*PyCAC3*、*PyCAC3L*、*PyDGATIL*)的表达量达到峰值,且它们在玉露香梨中的表达量相对更高。在本研究中,笔者发现,货架7 d后,对照组梨果皮蜡质合成代谢基因包括 *PyLACS9*、*PyKCR1*、*PyKCS20* 和 *PyCER1* 的表达量上调。*LACS9* 是脂肪酸长链脂酰基辅酶A合酶合成的关键基因,在油料作物提高种子含油量中发挥关键作用^[32]。此外,本研究中 *PyLACS9* 与果实 *L* 值呈显著正相关,因此认为它与玉露香梨果实的油腻化程度显著相关。结合 *PyLACS9* 在玉露香梨脂质代谢途径中发挥作用,推测 *PyLACS9* 可能通过调控长链脂酰辅酶A的合成而促进脂肪酸的降解,进而加速果皮油腻化相关组分的合成。

Jiang等^[19]研究表明,脂肪酸去饱和酶(FADs)在促进苹果皮油腻化的发生中发挥重要作用,其中 *MdFAD27* 和 *MdFAD28* 作为关键基因通过调控 FADs 的活性进而促进果皮油腻化。闫丹^[33] 在研究中报道, *MdFAD2* 基因的表达水平与苹果果皮油腻的蜡组分中的酯类物质含量呈显著正相关。在本研究中,笔者挖掘到玉露香梨中脂质合成代谢调控基因 *PyFAD2*。在脂质代谢通路途径中, *PyFAD2* 可能通过促进 FADs 的合成进而促进长链脂肪酸合成,为果皮油腻化组分的合成提供底物,因此推测该基因可作为调控玉露香梨果皮油腻化的候选基因进行深入研究。

3.2 1-MCP对蜡质合成代谢基因和乙烯合成基因的调控作用

根据 Li等^[34] 研究报道,1-MCP抑制了冷藏期间苹果果皮蜡合成代谢基因 *MdLACS1*、*MdCER6*、*MdCER4* 和 *MdWSD1* 的表达,导致果实角质层蜡中的醇类、脂肪酸类和酯类的含量降低。在本研究中,1-MCP抑制了玉露香梨 *PyLACS9*、*PyKCR1*、*PyKCS20* 和 *PyCER1* 基因的表达。结合玉露香梨外观可发现1-MCP处理延缓了果皮的油腻化进程。因此推测, *PyLACS9*、*PyKCR1*、*PyKCS20* 和 *PyCER1* 可能是调控玉露香梨果皮油腻化的关键基因。相关性分析结

果表明, *PyLACS9* 和果面亮度 *L* 值呈显著正相关,而 *L* 值与果皮油腻化密切相关。因此 *PyLACS9* 是调控玉露香梨果皮油腻化的关键基因,而1-MCP处理通过抑制其表达进而抑制果皮油腻化的发生。

此外,在本研究中1-MCP处理还显著抑制了整个货架期间梨果实的乙烯合成基因 *PyACO2* 和 *PyACS-1* 的表达。Li等^[18] 的研究结果表明,采用1-MCP处理玉露香梨果实,不仅显著抑制了果皮蜡质合成代谢基因和蜡质转运相关基因,包括 *PyLACS1*、*PyLACS6*、*PyKCS1*、*PyKCS2*、*PyKCS4*、*PyKCS10L*、*PyKCS11L*、*PyKCS20*、*PyFDH*、*PyCER10*、*PyKCR1*、*PyABCG11L*、*PyABCG12*、*PyABCG21L*、*PyLTPG1*、*PyLTP4*、*PyCAC3*、*PyCAC3L* 和 *PyDGATIL* 的上调表达,乙烯合成基因 *PyACO1* 和 *PyACS1* 显著下调表达,且上述基因的表达水平存在一定的相关性。在本研究中,相关性分析结果表明, *PyLACS9* 与 *PyACS-1* 呈负相关, *PyKCR1*、*PyCER1* 与 *PyACO2* 呈正相关, *PyPLDALPHA4* 与 *PyACS-1* 呈正相关, *PyFAD2* 与 *PyACO2*、*PyACS-1* 均呈负相关。由此推测,1-MCP还可能通过调控乙烯合成相关基因的表达水平进而影响果皮蜡质合成代谢基因的表达。

4 结 论

与对照相比,1-MCP处理对玉露香梨果皮蜡质合成代谢相关基因的影响主要富集到脂肪酸的延伸、合成和分解等次级脂质代谢途径。经1-MCP处理, *PyKCS20*、*PyCER1*、*PyLACS9*、*PyPLDALPHA4* 和 *PyFAD2* 下调表达,果面亮度 *L* 值较对照低。1-MCP可能通过影响上述基因的表达模式来调节果皮蜡质合成代谢途径,进而影响玉露香梨果皮油腻化水平。

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