

柑橘13个多胚品种同源四倍体高效发掘与分子鉴定

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摘要:【目的】基于柑橘珠心细胞存在自然加倍的特点, 实生播种发掘千山红蜜橘等13个品种的同源四倍体。【方法】种子催芽萌发后实生播种, 待幼苗长出3片以上真叶后, 依据形态特征初选法“观根辨叶看油胞”从实生苗中筛选疑似多倍体, 再通过流式细胞仪倍性分析与根尖染色体压片计数对疑似多倍体进一步鉴定倍性, 并通过SSR分子标记分析鉴定所获多倍体的遗传来源。【结果】基于形态初选, 分别从9个地方特色品种千山红蜜橘、八月橘、衢橘、早橘、扁平橘、瓠柑、狮头柑、冰糖橙、锦蜜冰糖橙和4个砧木品种磨坪香橙、日本香橙、积雀、油皮金柑的343、499、892、385、519、290、457、241、119、690、828、114、129株实生苗中发掘获得2、1、3、2、7、3、1、3、1、3、17、1、2株疑似多倍体; 用流式细胞仪对以上46株疑似多倍体进行倍性鉴定, 获得45株四倍体和1株衢橘六倍体, 采用根尖染色体计数法验证了上述结果; SSR分子鉴定表明, 13个品种的45株四倍体所扩增条带大小与其相应二倍体亲本完全一致, 推测其可能是由二倍体亲本珠心细胞自然加倍形成的同源四倍体。【结论】发掘的四倍体资源不仅丰富了我国柑橘多倍体类型, 而且为我国柑橘无核育种和矮化广适砧木选育奠定了珍贵的材料基础。

关键词: 柑橘; 多倍体; 流式细胞仪; SSR分子标记; 无核育种

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Efficient exploration and SSR identification of autotetraploids from the seedlings of thirteen apomictic *Citrus* genotypes

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Abstract: 【Objective】 *Citrus* is a crucial part of Chinese fruit crops. There are abundant citrus germplasm resources in China, but many excellent local varieties are gradually eliminated by the market due to the problem of numerous seeds within the fruit. The fruits of triploid plants are generally seedless because of their sterile male and female gametes. Therefore, triploid production is a promising strategy to breed seedless cultivars in citrus. Triploids can be obtained by interploidy crossing between diploids and tetraploids. However, the tetraploid germplasm is rare, which limits the application of this strategy. Exploration of tetraploids is an important prerequisite for triploid production with the aim to improve the seedy local cultivars in our country. For the rootstock improvement, tetraploid plants are also valuable resources because of their higher metabolite content, and better resistance than their diploid par-

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ents. In this study, we planned to explore tetraploid plants from 13 local cultivars in our country by using the traits of spontaneous doubling of the nucellar cells in polyembryonic citrus varieties. The exploration of tetraploids from the above 13 local cultivars will not only provide excellent tetraploid parents for the production of triploid plants, but also lay the foundation for the basic research about the effect of genome duplication on some important trait change, such as dwarfing, extensive adaptivity and higher medicinal value in tetraploids. **【Methods】** After the mature fruits were harvested, the seeds were extracted and the seed coats were peeled off, and then they were placed in a thermostat to accelerate germination. When the seeds germinated, they were sown in pots and cultivated in a plant growth chamber. After the seedlings grew up with three or more leaves, putative polyploids were screened according to the morphological feature showing lower height, shorter taproots, less lateral roots, thicker and rounder leaves and declined oil gland density. The ploidy levels of these putative polyploids were further confirmed by flow cytometric analysis and the observation on root tip chromosome numbers. After determination of the ploidy level, some morphological traits, including plant height, root length and diameter, lateral root number, stem diameter, leaf thickness and shape index of the tetraploids and their corresponding diploid parents were measured at the same developmental stage. SSR analysis was used to identify the genetic origin of the explored tetraploids with at least three pairs of SSR primers selected for each cultivar. **【Results】** The polyembryonic degree of seeds from each cultivar was firstly determined and it showed that the seeds of all 13 cultivars were polyembryonic. Among them, Qu tangerine had the highest number of embryos with an average of 9.4 embryos per seed and Bingtang sweet orange had the lowest number of embryos with an average of 2.2 embryos per seed. Based on the morphological trait screening, we identified 2, 1, 3, 2, 7, 3, 1, 3, 1, 3, 17, 1 and 2 putative polyploids respectively from 343, 499, 892, 385, 519, 290, 457, 241, 119, 690, 828, 114 and 129 seedlings of Qianshanhong tangerine, Bayue tangerine, Qu tangerine, Zao tangerine, Bianping tangerine, Ougan tangerine, Shitougan, Bingtang sweet orange, Jinmi sweet orange, Moping Xiangcheng, Japanese Xiangcheng, Zhique and Youpi kumquat. After further confirmation of ploidy levels concerning above putative tetraploids, we obtained 45 tetraploids and one hexaploid plant from Qu tangerine, with an average occurrence rate of 0.85%, among which the rate of Japanese xiangcheng was the highest with 2.05% and the rate of Bayue tangerine was the lowest with 0.20%. The exploration time from seed germination to obtaining tetraploid seedlings varied among cultivars, with the longest time (42 days) used in Youpi kumquat and the shortest time (23 days) in Shitougan. The morphological traits of tetraploids and their corresponding diploid seedlings from nine cultivars of Qianshanhong tangerine, Qu tangerine, Zao tangerine, Bianping tangerine, Ougan tangerine, Bingtang sweet orange, Moping Xiangcheng, Japanese Xiangcheng and Youpi kumquat were measured. For plant height, tap root length, lateral root numbers and leaf thickness, the tetraploid seedlings of seven cultivars showed significant differences with their diploid parents. For taproot and stem diameter, only the tetraploid seedlings explored from Bingtang sweet orange and Japanese Xiangcheng had significant difference with their diploid parents. For leaf shape index, the tetraploid seedlings from Bianping tangerine and Moping Xiangcheng exhibited significant differences with their diploid seedlings. In conclusion, most tetraploid seedlings of all nine cultivars showed lower plant height, shorter and thicker taproot, less lateral root number, thicker and rounder leaves than those of their diploid parents. These results provide supports for the screening of putative tetraploids based on morphological trait observation. For analyzing the genetic origin of the tetraploids obtained in this study, at least three SSR markers were used in each genotype. The results showed that the bands of all 45 tetraploids were identical with those of their corresponding diploids, indicating that all the 45 tetra-

ploids might originate from the spontaneous chromosome doubling of nucellar cells of their corresponding diploids. In addition, the bands of the hexaploid from Qu tangerine were also identical with their diploid parent. We speculated that it might derive from chromosome doubling of a triploid zygotic cell, which formed by selfing of a FDR-type $2n$ gamete with a normal n gamete, and both gametes were produced by Qu tangerine. 【Conclusion】 This study verified that morphological screening combined with flow cytometry ploidy determination and SSR analysis is an efficient approach to exploring polyploid seedlings from apomictic citrus. Using this method, 45 autotetraploid and one hexaploid plants were obtained from 13 apomictic citrus genotypes. These newly discovered tetraploids are potentially valuable for not only genetic improvement of some elite local citrus cultivars with seeds produced by triploids using interploidy hybridization, but also selection of the promising rootstocks with dwarf, multi-resistance and broad adaptability characteristics to improve the ability to resist various abiotic and biotic stresses.

Key words: *Citrus*; Polyploidy; Flow cytometry; Simple sequence repeat; Seedless breeding

柑橘是我国南方最重要的水果,在我国农村经济发展与乡村振兴中发挥了巨大的作用^[1]。我国柑橘栽培历史悠长,种质资源繁多,但许多优良的地方品种因种子多而逐渐被淘汰^[2]。无核是柑橘遗传改良的重要育种目标之一^[3],三倍体是天然的不育类型,创制三倍体是实现柑橘果实无核化的有效途径^[4]。利用二倍体与四倍体倍性杂交是培育柑橘三倍体的主要方法^[5],但我国一些地方特色品种四倍体资源仍较为缺乏,限制了该育种策略的应用。因此,发掘柑橘四倍体是利用倍性杂交策略改良我国地方特色品种种子多等性状的重要前提条件。此外,与二倍体相比,同源四倍体大部分表现为器官更大^[6]、代谢物含量更高^[7]、抗性更强^[8]和植株矮化或致矮^[9]等特点,发掘或创制柑橘砧木同源四倍体,对培育矮化、广适和多抗砧木品种具有重要的产业应用价值。

柑橘属及近缘属大多数品种均具有多胚现象^[10]。据报道,少数珠心细胞在发育过程中会发生自然加倍,形成染色体加倍的珠心胚萌发后进而形成同源四倍体(或称双二倍体),其基因型与二倍体完全一致^[11]。利用这一特性,梁武军等^[9,12]通过露地实生播种结合叶片形态观察发掘获得了枳、香橙、早金甜橙等20余个砧木和柑橘接穗品种的同源四倍体。但是,种子露地播种易受季节和天气等环境因素影响,存在萌发率低且初选准确率不高的缺点。针对该问题,周锐等^[13]对上述技术流程进行了改良,提出了“观根辨叶看油胞”发掘四倍体的方法,该方法采用种子催芽后生长室播种代替露地播种,综合叶片、根系和油胞等多组织形态特征(如四倍体相较二倍体根短粗且侧根少、叶片厚、叶形指

数小、油胞密度低等)筛选疑似多倍体,极大提高了发掘效率。基于该方法,谢善鹏等^[14]以常山胡柚等6个柑橘地方特色品种为材料,发掘获得了53株四倍体新种质,准确率46.7%,从播种到获得四倍体耗时平均38 d。张成磊等^[15]利用该方法从1289株山金柑实生后代中发掘同源四倍体8株,准确率100%,证明了“观根辨叶看油胞”结合流式细胞仪倍性鉴定是获得柑橘多胚品种同源四倍体的便捷高效的策略。

千山红蜜橘、八月橘、衢橘、早橘、扁平橘、瓯柑、冰糖橙和油皮金柑是我国陕西、广东、浙江和湖南等地的优良地方特色品种,但均存在种子较多和食用不便的问题;香橙、枳雀和狮头柑除了作为优良的砧木资源外,还具有一定的药用价值。发掘上述材料的多倍体新种质,不仅能为柑橘三倍体无核种质创制提供优良的四倍体育种亲本,而且对培育矮化、适应性强的四倍体砧木和药用价值高的柑橘资源具有重要研究意义和实践价值。

1 材料和方法

1.1 试验材料

9个地方特色品种:千山红蜜橘(*Citrus reticulata* Blanco)、八月橘(*C. reticulata* Blanco)、衢橘(*C. reticulata* Blanco)、早橘(*C. reticulata* Blanco)、扁平橘(*C. Depressa* Hayata)、瓯柑(*C. reticulata* Sua-vissima)、冰糖橙(*C. sinensis* L. Osbeck)、锦蜜冰糖橙(*C. sinensis* L. Osbeck)和油皮金柑(*Fortunella japonica* Swingle);4个砧木品种:磨坪香橙(*C. junos* Siebold ex Tanaka)、日本香橙(*C. junos* Siebold ex Tanaka)、枳雀(*C. wilsonii* Tanaka)和狮头柑(*C. spe-*

ciosa Hort. ex Tseng)。

八月橘采自广东省深圳市;冰糖橙、锦蜜冰糖橙采自湖南省郴州市;狮头柑、枳雀、日本香橙和千山红蜜橘采自陕西省城固县;磨坪香橙采自湖北省秭归县;衢橘、早橘和扁平橘采自浙江省衢州市;瓯柑采自浙江省温州市;油皮金柑采自广西融安县。

1.2 实生播种及初选疑似多倍体

种子实生播种参考谢善鹏等^[14]的方法,实生幼苗形态初选疑似多倍体参考周锐等^[13]的方法。从成熟果实剥取种子后,用 $1 \text{ mol} \cdot \text{L}^{-1}$ KOH 溶液浸泡 3~5 min 溶解果胶,用清水搓洗干净后剥去种皮,置于垫有湿纱布的玻璃皿于 $28 \text{ }^{\circ}\text{C}$ 恒温暗催芽 3~5 d;待胚根长至 0.5~1.0 cm 时,播种于塑料钵并覆膜保湿,置于生长室(温度 $25 \pm 1 \text{ }^{\circ}\text{C}$,光照 16 h)培养。待幼苗长出 3 片以上真叶后,依据形态特征初选法“观根辨叶看油胞”从实生苗中筛选出疑似多倍体。

1.3 倍性分析与鉴定

流式细胞仪(Cyflow space, Sysmex, Japan)倍性分析参考解凯东等^[16]的方法。用一步法试剂盒(Sysmex, Germany)对样品进行裂解和染色后上机

检测,以二倍体($2n=2x=18$)植株叶片(荧光强度 $X \approx 50$)为对照,分析待测样品倍性,若待测样品为三倍体($2n=3x=27$),则其荧光强度 $X \approx 75$;若待测样品为四倍体($2n=4x=36$),则其荧光强度则为 $X \approx 100$ 。植株根尖染色体压片计数参考 Xia 等^[17]的方法。植株倍性确定后,统计各个品种四倍体的自然发生频率(四倍体株数/群体株数)。

1.4 四倍体幼苗形态指标测定

参考谢善鹏等^[14]的方法对相同发育时期的四倍体及二倍体实生苗形态指标进行测定。形态指标主要包括植株高度、主根长度、侧根数目、根粗(根茎分界处向下 1 cm 处的根直径)、茎粗(根茎分界处向上 1 cm 处的茎直径)、叶片厚度和叶形指数(叶片长度/叶片宽度)。每项指标均随机取样测定 3 个生物学重复,每个品种二倍体及四倍体实生苗各测量 9 株。用 Excel 2016 对数据进行处理,并用 t -test($p < 0.05$)对数据进行显著性分析。

1.5 植物基因组 DNA 提取和 SSR 分子鉴定

参考 Cheng 等^[18]的方法提取基因组 DNA,SSR 分子标记参考谢善鹏等^[14]的方法。SSR 引物见表 1,由北京擎科生物科技股份有限公司武汉分公司合成。PCR 扩增体系为 $10 \mu\text{L}: 2 \times \text{Rapid Taq Master}$

表 1 SSR 引物序列
Table 1 Sequence of SSR primers

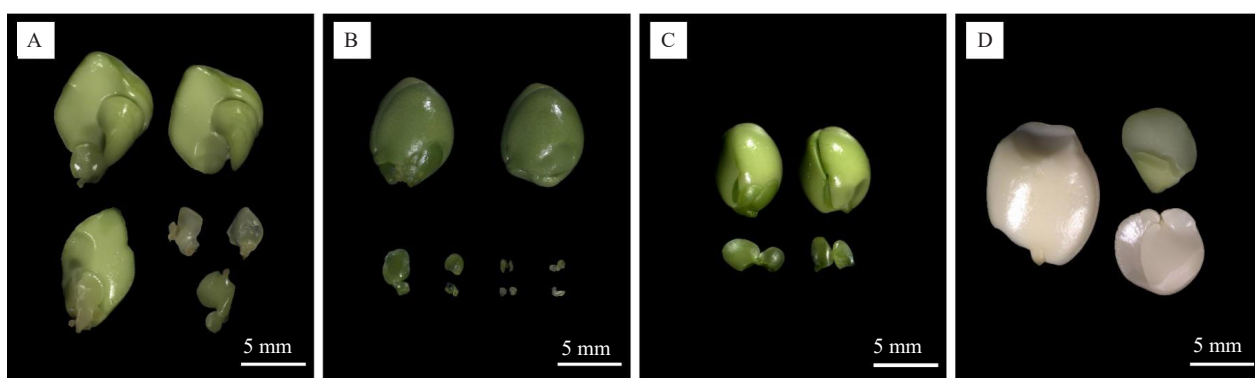
引物名称 Primer name	引物序列 Primer sequence	文献 References
CX296884	F: GCTCCTCGAATGAGAATGAAATGA; R: TGGTTGTGCGAAAATGAAGAGATA	[19]
CD575830	F: TACCATGTGCCCTTTCTGCTATTT; R: AGCTATGGCTTTGGTTGAGTTCTG	[19]
M1H3Si6130	F: ATAAGGCCCGATATGGGAAG; R: GCATGAAATGATTGAGCCAG	[19]
CV710821	F: GATACAAATTAGCATTGATTGAATGGA; R: ATCGGGACTCGCATTAGGGT	[19]
mCrCIR02D09	F: AATGATGAGGGTAAAGATG; R: ACCCATCACAAAACAGA	[20]
mCrCIR05A05	F: ATACCTGTGAGCGTGAG; R: CCTCTTCCCTTCCATT	[20]
mCrCIR07D06	F: CCTTTTCACAGTTTGCTAT; R: TCAATTCCTCTAGTGTGTGT	[20]
MEST1	F: CAAGCCTCTCTTTTAGTCCCA; R: AGTTCTTTGGTGCTTCAGGC	[21]
mCrCIR01F08a	F: ATGAGCTAAAGAGAAGAGG; R: GGACTCAACACAACACAA	[22]
mCrCIR06B07	F: CGGAACAACATAAAACAAT; R: TGGGCTTGATAGACAGTTA	[22]
Ci01C07	F: GTCACACTCTCGCTCTTG; R: TTGCTAGCTGCTTTAACTTT	[22]
Ma2-1556	F: TTGCATGGCTTCATGTTAGG; R: TAAGAACAACCCTGACCCA	[23]
Ma3-59	F: TTACAGTCACAGCAGCCTGG; R: AGAAGAGGACGAGGGAGAGG	[23]
Csin0341	F: ACAGTGTGTCCGCATGAAAA; R: ATTCCTTACAACCGTGCT	[23]
Ma3-153	F: CTGTTGCTGCTTTGGATCA; R: GTTCCGGATTGAACCATGTC	[23]
Csin0153	F: GTCTGGGTGAGTGTGGAGT; R: CGAAAAAGACAGCCAAATCC	[23]
Ma2-1480	F: CAATCACAGGAGCGACTTCA; R: CTCAATTCAGCAAACCGACA	[23]
CAT01	F: GCTTTCGATCCCTCCACATA; R: GATCCCTACAATCCTTGGTCC	[24]
cAGG9	F: AATGCTGAAGATAATCCGCG; R: TGCCTTGCTCTCCACTCC	[25]

Mix 5 μL , ddH₂O 3.5 μL , DNA 1 μL , Primer F、R 各 0.25 μL (10 $\mu\text{mol}\cdot\text{L}^{-1}$)。PCR 反应在 ProFlex PCR 仪 (ABI, USA) 进行, 扩增程序: 95 $^{\circ}\text{C}$ 预变性 3 min, 95 $^{\circ}\text{C}$ 变性 30 s, 55 $^{\circ}\text{C}$ 退火 30 s, 72 $^{\circ}\text{C}$ 延伸 10 s, 35 个循环, 72 $^{\circ}\text{C}$ 延伸 5 min, 4 $^{\circ}\text{C}$ 保存。PCR 扩增产物由全自动毛细管电泳 (QIAxcel Advanced, Germany) 分离, 成像结果由仪器自带软件 QIAxcel ScreenGel 自动生成。

2 结果与分析

2.1 13 个柑橘品种均为多胚品种

对千山红蜜橘、八月橘、衢橘、早橘、扁平橘、瓯柑、狮头柑、冰糖橙、锦蜜冰糖橙、磨坪香橙、日本香橙、枳雀和油皮金柑共 13 个品种的种子胚数进行调查 (图 1), 每个品种至少统计 20 粒种子。结果表明 (表 2), 上述 13 个品种均为多胚品种, 种子多胚程度



A. 早橘; B. 油皮金柑; C. 扁平橘; D. 日本香橙。

A. Zao tangerine; B. Youpi kumquat; C. Bianping tangerine; D. Japanese Xiangcheng.

图 1 4 个柑橘品种种子多胚程度观察

Fig. 1 Polyembryonic degree in the seeds from four citrus cultivars

表 2 千山红蜜橘等 13 个多胚品种四倍体实生发掘情况

Table 2 Exploration of the tetraploid from 13 citrus polyembryonic cultivars

品种 Cultivar	单果种子数 No. of seeds per fruit	种子胚数 No. of embryos	催芽种子数 No. of seeds	萌发数 No. of seedlings	四倍体数 No. of tetraploid seedlings	群体发生率 Population incidence/%	耗时 Time/d
千山红蜜橘 Qianshanhong tangerine	6.4±3.5	2.8±1.1	253	343	2	0.58	39
八月橘 Bayue tangerine	11.5±3.0	3.7±0.9	688	499	1	0.20	38
衢橘 Qu tangerine	13.7±4.0	9.4±3.5	668	892	2	0.22	23
早橘 Zao tangerine	1.9±2.6	4.5±1.8	178	385	2	0.52	30
扁平橘 Bianping tangerine	6.7±3.0	4.4±1.8	709	519	7	1.35	38
瓯柑 Ougan tangerine	5.2±4.3	2.5±1.3	271	290	3	1.03	34
狮头柑 Shitougan	21.4±5.3	7.8±2.5	103	457	1	0.22	23
冰糖橙 Bingtang sweet orange	1.8±1.6	2.2±1.0	163	241	3	1.24	27
锦蜜冰糖橙 Jinmi sweet orange	1.8±1.4	4.6±1.1	81	119	1	0.84	31
磨坪香橙 Moping Xiangcheng	28.9±3.8	4.0±2.7	617	690	3	0.43	34
日本香橙 Japanese Xiangcheng	26.6±6.0	2.7±1.8	846	828	17	2.05	29
枳雀 Zhique	62.6±8.2	3.3±1.0	119	114	1	0.88	31
油皮金柑 Youpi kumquat	4.2±1.3	7.2±2.9	120	129	2	1.55	42

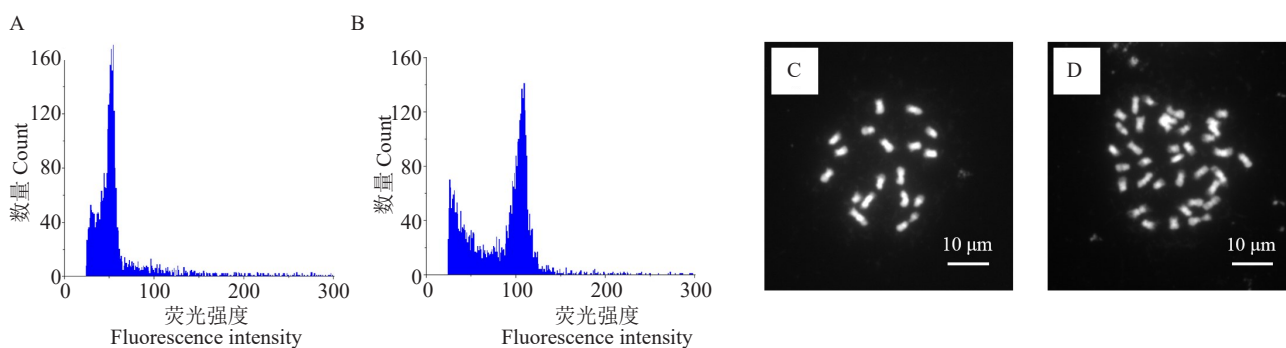
由低到高依次为冰糖橙、瓯柑、日本香橙、千山红蜜橘、枳雀、八月橘、磨坪香橙、扁平橘、早橘、锦蜜冰糖橙、油皮金柑、狮头柑和衢橘。其中, 衢橘胚数最高, 平均每粒种子 9.4 个胚; 冰糖橙胚数最少, 平均每粒种子 2.2 个胚。

2.2 实生发掘 13 个柑橘品种同源四倍体 45 株

依据“观根辨叶看油胞”形态学筛选法, 分别从千山红蜜橘、八月橘、衢橘、早橘、扁平橘、瓯柑、狮头柑、冰糖橙、锦蜜冰糖橙、磨坪香橙、日本香橙、枳雀和油皮金柑 13 个品种的 343、499、892、385、519、

290、457、241、119、690、828、114、129株实生苗筛选获得2、1、3、2、7、3、1、3、1、3、17、1、2株疑似多倍体。用流式细胞仪对上述46株疑似多倍体进行倍性鉴定(图2),获得45株四倍体(千山红蜜橘2株、八月橘1株、衢橘2株、早橘2株、扁平橘7株、瓯柑3株、狮头柑1株、冰糖橙3株、锦蜜冰糖橙1株、磨坪香橙3株、日本香橙17株、枳雀1株、油皮金柑2株)和1株六倍体(衢橘)。进一步随机挑选日本香橙的二倍体和四倍体实生幼苗各3株,利用根尖染色体

计数对其染色体数目进行鉴定,结果表明,流式细胞仪测得的四倍体染色体为36条,二倍体为18条,验证了流式细胞仪倍性分析的准确性。不同品种四倍体的群体发生率不同,发生率由低到高依次为八月橘、衢橘、狮头柑、磨坪香橙、早橘、千山红蜜橘、锦蜜冰糖橙、枳雀、瓯柑、冰糖橙、扁平橘、油皮金柑、日本香橙。其中,日本香橙群体发生率最高,为2.05%;八月橘群体发生率最低,为0.20%。从播种至获得四倍体植株,不同品种耗时23~42 d不等(表2)。



A、B. 流式细胞仪倍性分析实生幼苗倍性(A. 二倍体, 荧光强度 $X \approx 50$; B. 四倍体, 荧光强度 $X \approx 100$); C、D. 根尖染色体压片计数鉴定实生幼苗倍性(C. 二倍体, $2n=2x=18$; D. 四倍体, $2n=4x=36$)。

A, B. Ploidy determination of the seedlings by flow cytometry analysis (A. Diploid, fluorescence intensity $X \approx 50$; B. Tetraploid, fluorescence intensity $X \approx 100$); C, D. Chromosome number analysis by root tip chromosome counting technique (C. Diploid, $2n=2x=18$; D. Tetraploid, $2n=4x=36$).

图2 流式细胞仪倍性分析及根尖染色体压片计数鉴定日本香橙实生幼苗倍性

Fig. 2 Ploidy determination of the seedlings from Japanese xiangcheng by flow cytometry analysis and root tip chromosome counting technique

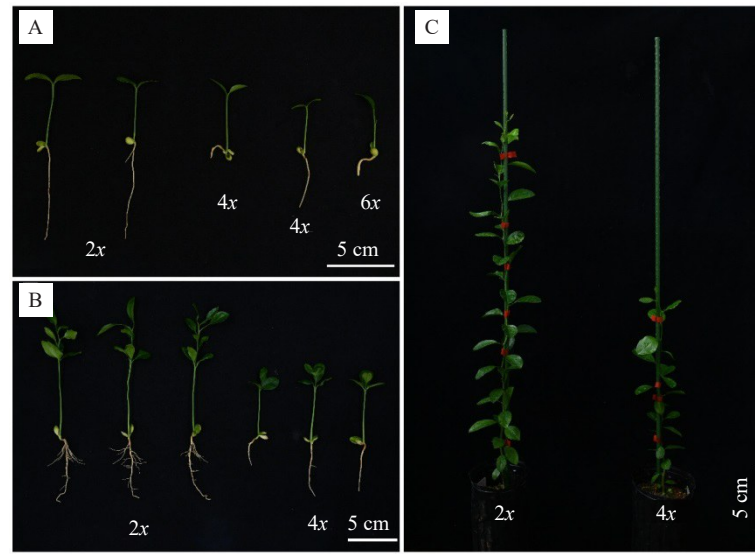
2.3 9个品种的四倍体与二倍体幼苗根、茎和叶片形态特征差异明显

对千山红蜜橘、衢橘、早橘、扁平橘、瓯柑、冰糖橙、磨坪香橙、日本香橙和油皮金柑9个品种发掘获得的四倍体及相应二倍体亲本实生幼苗的株高、主根长度、侧根数目、根粗、茎粗、叶片厚度、叶片长度和宽度等形态指标进行测定,以评价四倍体及其二倍体亲本实生幼苗的形态差异(图3)。结果(表3)表明,除冰糖橙和油皮金柑外,其余7个品种的四倍体幼苗株高显著低于其对应二倍体亲本;主根长度方面,除瓯柑和冰糖橙外,其余7个品种的四倍体幼苗主根长度显著低于其对应二倍体亲本;对于侧根数和根粗,除早橘和冰糖橙外,其余7个品种的四倍体幼苗侧根数显著低于其二倍体亲本,且仅有冰糖橙四倍体幼苗根显著粗于其二倍体亲本,其余品种根粗差异不明显;而在茎粗方面,大多数品种的四倍体幼苗与其二倍体亲本差异不明显,仅有日本香橙的四倍体茎显著粗于其二倍体亲本;在叶片厚度和

叶形指数方面,除磨坪香橙和日本香橙外,其余7个品种的四倍体幼苗叶片厚度显著大于其二倍体亲本,扁平橘和磨坪香橙的四倍体幼苗的叶形指数显著小于其二倍体亲本,其余品种差异不显著。总的来说,大多数品种的四倍体幼苗与其对应二倍体相比,植株一般表现为株高显著降低,主根变短粗且侧根减少,叶片增厚、叶形指数变小等特点。

2.4 SSR分子鉴定

用19对SSR引物对从13个品种发掘获得的四倍体和六倍体植株进行SSR分子鉴定,每个品种至少选择3对多态性较好的引物进行鉴定(图4)。结果表明,13个品种的四倍体所扩增条带的大小与其相应二倍体亲本所扩增条带大小完全一致,推测其可能由二倍体亲本珠心细胞自然加倍形成的同源四倍体;衢橘六倍体条带也与其二倍体亲本基本一致,推测其可能由衢橘形成的 $2n$ 配子与正常 n 配子自交形成三倍体合子后,再经染色体自然加倍形成。



A. 衢橘 23 d 幼苗形态; B. 日本香橙 29 d 幼苗形态; C. 八月橘在温室生长 180 d 左右的幼苗形态。

A. The seedling morphology of Qu tangerine growing for 23 d; B. Japanese Xiangcheng growing for 29 d; C. Bueyue tangerine growing for 180 d.

图 3 不同品种四倍体或六倍体与其二倍体亲本实生幼苗形态比较

Fig. 3 Morphological characteristics of the tetraploid and hexaploid seedlings comparing with their corresponding diploid parents

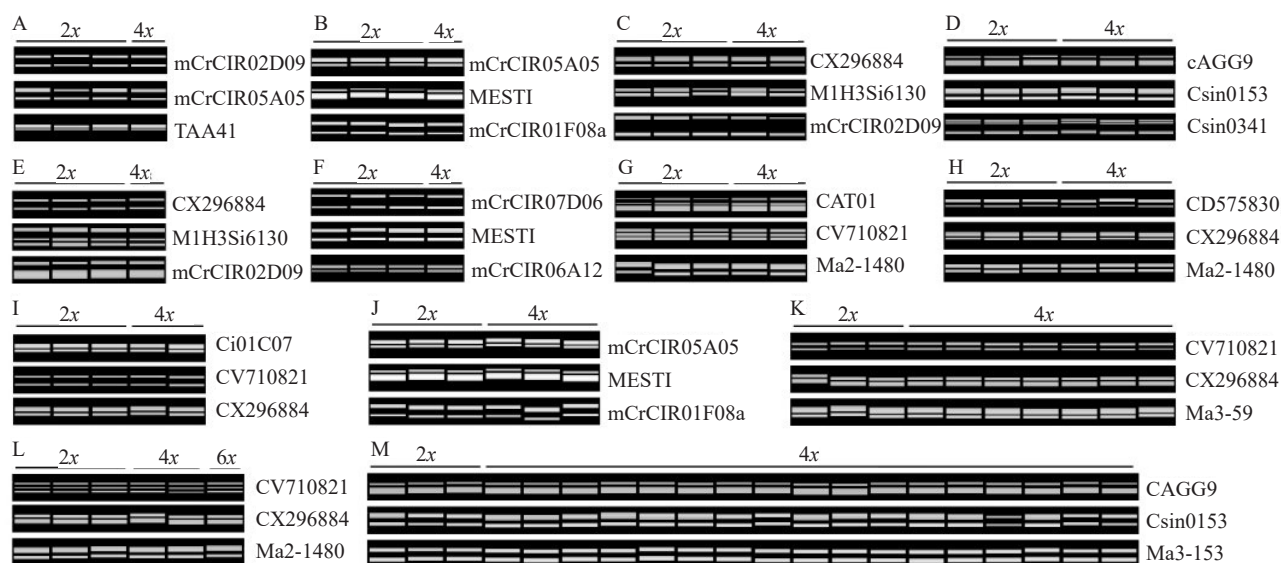
表 3 千山红蜜橘等 9 个品种四倍体及其相应二倍体植株幼苗形态特征比较

Table 3 Comparison of morphological traits of the root, stem and leaf between the seedlings of tetraploids from nine polyembryonic cultivars and their corresponding diploid parents

品种 Cultivar	倍性 Ploidy	株高 Plant height/ mm	主根长 Root length/ mm	侧根数 No. lateral roots	根粗 Root diameter/ mm	茎粗 Stem diameter/ mm	叶片厚度 Leaf thickness/ mm	叶形指数 Leaf shape index
千山红蜜橘 Qianshanhong tangerine	2x	69.34±7.36	80.53±9.90	11.89±2.62	1.01±0.08	1.21±0.31	0.14±0.01	2.12±0.24
	4x	33.07±13.97**	42.00±1.61**	1.50±2.12**	0.86±0.11	1.02±0.13	0.16±0.01**	1.93±0.31
衢橘 Qu tangerine	2x	55.25±7.83	51.11±17.44	7.11±2.52	0.93±0.28	1.00±0.16	0.13±0.01	1.34±0.27
	4x	26.79±2.73**	19.31±3.78*	0.00±0.00**	0.75±0.02	0.81±0.02	0.15±0.01*	1.57±0.26
早橘 Zao tangerine	2x	52.82±4.33	71.64±12.02	1.33±1.41	0.86±0.07	0.94±0.12	0.14±0.00	1.48±0.14
	4x	37.76±7.82**	30.10±11.20**	0.33±0.58	1.08±0.48	0.97±0.08	0.17±0.03**	1.62±0.06
扁平橘 Bianping tangerine	2x	64.59±7.02	92.26±14.82	11.67±3.08	0.93±0.11	1.02±0.10	0.12±0.01	2.16±0.33
	4x	35.64±2.27**	18.19±10.89**	1.00±0.00**	0.92±0.01	1.02±0.06	0.10±0.02*	1.40±0.08*
瓯柑 Ougan tangerine	2x	83.78±9.07	122.51±9.07	18.56±2.24	1.53±0.17	1.54±0.11	0.14±0.01	1.75±0.20
	4x	64.34±7.08**	106.15±17.30	12.00±1.73**	1.52±0.31	1.54±0.20	0.17±0.00**	1.65±0.22
冰糖橙 Bingtang sweet orange	2x	78.75±9.37	77.06±26.02	9.11±2.03	1.24±0.13	1.36±0.13	0.20±0.01	1.50±0.22
	4x	74.05±9.87	75.79±19.58	7.00±2.65	1.68±0.08**	1.53±0.27	0.25±0.01**	1.38±0.08
磨坪香橙 Moping Xiangcheng	2x	74.35±6.54	95.96±12.01	9.44±3.94	1.23±0.13	1.20±0.16	0.13±0.01	1.84±0.24
	4x	54.50±15.07**	35.89±12.16**	1.84±3.00**	1.15±0.16	1.38±0.28	0.21±0.24	1.45±0.20**
日本香橙 Japanese Xiangcheng	2x	114.08±13.78	85.26±8.20	13.56±4.45	1.76±0.23	1.64±0.12	0.13±0.01	1.71±0.31
	4x	80.27±17.38**	56.07±15.23**	4.00±3.80**	1.73±0.43	1.81±0.22*	0.14±0.02	1.50±0.33
油皮金柑 Youpi kumquat	2x	62.41±6.48	72.70±6.03	7.00±1.58	1.23±0.16	1.03±0.08	0.11±0.01	1.68±0.35
	4x	51.05±11.95	39.05±11.95**	1.50±2.12**	1.42±0.16	1.04±0.02	0.15±0.02**	1.76±0.10

注:表中数据(平均值±标准差)后面*和**分别表示在 $p < 0.05$ 和 $p < 0.01$ 水平上差异显著。

Note: The values (mean ± standard deviation) with * ($p < 0.05$) and ** ($p < 0.01$) indicate the significant difference by t -test.



A. 枳雀; B. 锦蜜冰糖橙; C. 千山红蜜橘; D. 磨坪香橙; E. 八月橘; F. 狮头柑; G. 早橘; H. 瓯柑; I. 油皮金柑; J. 冰糖橙; K. 扁平橘; L. 衢橘; M. 日本香橙。

A. Zhique; B. Jinmi sweet orange; C. Qianshanhong tangerine; D. Moping Xiangcheng; E. Bayue tangerine; F. Shitougan; G. Zao tangerine; H. Ougan tangerine; I. Youpi kumquat; J. Bingtang sweet orange; K. Bianping tangerine; L. Qu tangerine; M. Japanese Xiangcheng.

图 4 千山红蜜橘等 13 个品种发掘的四倍体和六倍体幼苗 SSR 分子鉴定

Fig. 4 SSR profiles of the tetraploid and hexaploid plants with their corresponding diploid parents

3 讨 论

除枸橼类、柚类等以外,柑橘绝大多数品种的种子都具有多胚现象^[10],即一粒种子萌发会产生多株幼苗,其中可能既有合子苗(有性苗)又有珠心苗(无性苗)。一方面,因合子胚在与珠心胚发育中营养竞争处于劣势地位^[26],常导致合子胚败育,较少部分为有性杂种,很大程度上降低了杂交育种的效率;另一方面,由于珠心苗与母本遗传组成完全相同,属于无性后代,播种后的植株能保持母本优良性状,便于固定杂种优势^[27],具有重要的育种应用价值。柑橘多胚品种珠心细胞自然加倍的现象被报道以来^[5,28-30],大量多胚品种的同源四倍体资源被发掘^[9,11-15]。特别是将植株外部特征(包括株高度、根长度和粗度、叶片颜色和厚度、油胞密度等指标)作为形态标记用于初选多倍体,使得柑橘多倍体发掘技术趋于完善,效率也得到很大提升,准确率超过 80%^[13]。本研究基于课题组前期建立的“观根辨叶看油胞”形态初选结合流式细胞仪倍性快速鉴定的方法,从千山红蜜橘、八月橘等 13 个柑橘多胚品种的 5506 株实生苗中发掘获得了四倍体 45 株和六倍体 1 株,不仅验证了利用“观根辨叶看油胞”形态观察初选柑橘多倍体的高效性,还在一定程度上丰富了柑橘多倍体种质资源,

为未来更好地对我国柑橘地方特色资源进行无核改良或培育矮化、广适砧木奠定了宝贵的材料基础。

用 SSR 分子标记鉴定所获多倍体的遗传来源,13 个品种的 45 株四倍体的扩增条带的大小与其相应二倍体亲本的条带完全一致,表明其可能由二倍体亲本珠心细胞自然加倍形成的同源四倍体,这与前人的研究结果基本一致^[11-15]。而从衢橘中发掘获得的六倍体条带与其二倍体亲本条带也完全一致,未发现新的条带,推测其可能由衢橘产生的杂合型 $2n$ 配子(第一次减数分裂异常产生,多数位点基因型与二倍体亲本一致)与其正常的 n 配子自交形成三倍体合子后再加倍形成,该假设后期可利用近着丝粒标记^[17]或随机筛选更多的标记^[31]对该六倍体进行基因分型以确定其遗传来源。与四倍体相比,关于柑橘六倍体的报道相对较少。梁武军等^[32]通过三倍体葡萄柚胚培养获得了包括六倍体在内的一些不同倍性植株和疑似非整倍体植株。Guo 等^[33]通过细胞融合技术,创制获得了锦橙(二倍体)+HR(哈姆林甜橙+粗柠檬异源四倍体体细胞杂种)的异源六倍体。与其他倍性植株相比,六倍体植株生长速度极慢、植株极度矮小、叶片更小且厚、颜色更深,生产应用价值还需进一步评价。但是,作为基础研究材料,六倍体植株对开展柑橘倍性效应等相关基础研究具

有重要价值。

4 结 论

通过“观根辨叶看油胞”形态学初选结合流式细胞仪倍性鉴定,快速高效发掘了千山红蜜橘等13个品种的45株同源四倍体以及1株衢橘六倍体,遗传鉴定显示所有四倍体均为其对应二倍体亲本的珠心细胞自然加倍形成的同源四倍体。这些四倍体新种质不仅为我国柑橘地方特色品种的无核改良提供了宝贵的四倍体育种亲本,同时也为选育矮化、多抗和适应性强的四倍体砧木奠定了丰富的材料基础。

致谢:本研究所用的磨坪香橙和油皮金柑果实分别由国家现代农业(柑橘)产业技术体系三峡库区脐橙综合试验站曹立新站长和柳州综合试验站陆文科站长提供,特此感谢!

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