

方位及冠层对核桃内果皮木质素形成 及相关酶活性的影响

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摘要:【目的】明确核桃果实发育过程中不同光照强度对内果皮中木质素、相关酶活性影响的动态变化规律。【方法】以核桃品种‘新新2号’为试验材料,研究了树冠不同方位、不同冠层对内果皮中木质素含量、相关酶活性的变化规律,并进行了相关性分析。【结果】在不同方位、不同冠层上,木质素含量随着核桃果实的生长发育呈持续增加趋势;PAL、POD酶活性强弱的排序依次为:南面上层>南面中层>南面下层>北面上层>北面中层>北面下层>中部上层>中部中层>中部下层;PPO酶活性强弱的排序依次为:北面下层>北面中层>北面上层>中部下层>中部中层>中部上层>南面下层>南面中层>南面上层。相关性分析表明,木质素含量与POD酶活性呈极显著正相关($p < 0.01, r = 0.808^{**}$),与PPO酶活性呈负相关。【结论】树冠南面和上层接受的光照强度最大,木质素含量最高,PAL、POD酶活性最强;PPO酶活性最低;木质素含量与POD酶活性呈极显著正相关($p < 0.01, r = 0.808^{**}$),表明光照强度诱导了核桃果实内果皮中木质素的积累及相关酶活性的增强。

关键词:核桃;内果皮;木质素;酶活性;方位;冠层

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Effects of canopy exposure and layers on lignin formation and related enzyme activities in endocarp of walnut

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Abstract: 【Objective】Walnut (*Juglans regia* L.) is one of the most important traditional economic forest trees in China. It is cultivated in 1 046 counties and cities across the country, and its cultivated area has increased by 10% per year in recent years. Walnut nuts are composed of hard shells and seed kernels. The seed kernels are rich in nutrients, and have the functions of brain-enhancing, nourishing blood, warming the lungs and invigorating the intestines, moistening and drying phlegm, etc., which are deeply loved by people. In the past times, the lignified hard shells were mostly abandoned and attention was rarely received. As an important part of walnut nuts, hard shell plays an important role in nut growth, development, maturation, rinsing, transportation and storage, and activated carbon produced from hard shells can effectively absorb mercury, acid dye, phenol, etc. in industrial wastewater. Therefore, it is of great significance to study the formation mechanism and influencing factors of walnut nut hard shell. In recent years, with the continuous expansion of walnut growing area, the early-bearing walnuts with short growth cycle, high yield and good all-in-one have been vigorously promoted. However,

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in the actual production, the early-bearing walnut orchard has a common problem that affects the appearance quality of walnut nuts, such as “white face” and “Lu Ren” (Exposed kernel). In order to improve this phenomenon, accelerate the development of the walnut industry and improve the economic value of the walnut market, the growth and development of endocarp and the main factors affecting its growth and development were studied to explore the dynamic changes of the effects of light intensities on the activity of lignin and related enzymes in endocarp during the development of walnut fruit.【Methods】‘Xinxin No. 2’ cultivar with a planting space of 3 m × 6 m was used as a material in the walnut seedling demonstration garden of Xinjiang Academy of Forestry in Wensu county, Akesu region. Six sample trees with relatively consistent vigor were chosen and nuts were collected from the same part of each tree and mixed, and the mixed nut sample was divided into three parts as three replicates. During sampling, the sample tree was divided into different parts. Fruit samples were collected from the lower layer (less than 2.5 m from the base of the trunk), the middle layer (2.51 to 5 m from the base of the trunk), and the upper layer (more than 5m from the base of the trunk). On the horizontal distribution, fruit samples were picked in the middle of the tree (within 1.5 m from the central trunk), south and north (more than 1.5 m from the central trunk); from June 5, 2019 to July 26, 2019, at 7 d intervals. Sampling was performed once for a total of 7 times. After each picking, the fruits were classified and bagged according to different parts, and brought back to the laboratory for storage at -40 °C in a low-temperature refrigerator for the determination of various indicators. To observe the continuous deposition process of endodermal lignin, the content of lignin and the activities of related enzymes (PAL, POD, PPO) were determined. According to the changes of lignin deposition, the trend of lignin content and the change of related enzyme activities were analyzed in relation to the different positions of the canopy. Effects of light intensities on endodermal lignin deposition, lignin content, and changes in related enzyme activities were studied, and correlation analysis was performed.【Results】Lignin deposition began to accumulate from both ends of the endocarp to the middle, and the color changed from cherry red to purplish red, and the thickness and hardness of the endocarp increased with the increase of walnut fruit. The lignin content continued to increase with the development of walnut fruits. On different exposures and in different canopy layers, the order of the lignin content was as follows: south & upper layer > south & middle layer > south & lower layer > north & upper layer > north & middle layer > north & lower layer > middle & upper layer > middle & middle layer > middle & lower layer. On different exposures and in different canopy layers, the order of PAL and POD enzyme activity was as follows: south & upper layer > south & middle layer > south & lower layer > north & upper layer > north & middle layer > north & lower layer > middle & upper layer > middle & middle layer > lower & middle layer; The order of PPO enzyme activity was: north & lower layer > north & middle layer > north & upper layer > middle & lower layer > middle & middle layer > middle & upper layer > south & lower layer > south & middle layer > south & upper layer. Correlation analysis showed that lignin was significantly positively correlated with POD enzyme activity ($p < 0.01$, $r = 0.808^{**}$), and negatively correlated with PPO enzyme activity.【Conclusion】The south-exposed and upper canopy layer received the highest light intensity, and the nuts had the highest lignin content, the strongest PAL and POD enzyme activities and the lowest PPO enzyme activity. The lignin content was extremely significantly positively related to the POD enzyme activity ($p < 0.01$, $r = 0.808^{**}$), indicating that light intensity induced the lignin accumulation and enhanced the related enzyme activities in the endocarp of walnut fruits.

Key words: Walnut; Endocarp; Lignin; Enzymatic activity; Exposure; Canopy

光合作用对果树的生长发育过程具有重要的影响作用^[1-2],果树通过光合作用利用太阳能合成自身所需的碳水化合物^[3],而且光合同化产物也是果树产量与品质形成的物质基础^[4-5],作物干物质积累的90%~95%来源于光合作用^[6]。光照强度是影响光合作用的主要环境因子^[7-8],光照强度的变化会引起植物光合生理发生改变^[9-12]。近年来,随着新疆特色林果产业的迅速发展,核桃已成为新疆阿克苏地区覆盖面最大、产业带动性最强、群众受益面最广的绿色支柱产业。由于核桃种植面积不断扩大,生长周期短、产量高、可取全仁的早实核桃得到了大力推广,其中核桃‘新新2号’为新疆各核桃产区主栽品种之一,其产量虽常年较高,但其坚果外观品质受“白脸”、“露仁”等现象的影响,导致其在脱青皮、清洗、干燥等初加工过程中,核桃仁被污染率高,核桃商品化率降低,经济产值下降。

核桃坚果由内果皮(硬壳)及种仁组成,其内果皮中木质素的含量及组成是影响核桃坚果外观品质的主要因素。李夕勃^[13]的研究表明,同一品种不同部位的核桃内果皮中木质素含量和相关酶活性存在差异。宁万军等^[14]对两个品种的核桃进行6种元素肥的处理后,结果发现降低了其露仁率。靳万鑫等^[15]研究发现麻核桃中发育正常的内果皮其木质素远高于发育不良的内果皮,而纤维素低于发育不良的内果皮。鱼尚奇等^[16]对纸皮核桃硬化期的内果皮进行转录组测序,发现筛选出的候选基因参与了核桃内果皮硬化的生理生化过程。目前相关学者主要在核桃内果皮发育与木质素代谢的关系^[13,17-18]、核桃坚果内果皮发育的影响因子^[14,19]、不同发育程度与核桃内果皮主要成分的关系^[15]、木质素代谢过程中相关基因克隆与表达分析^[16,20-22]等方面进行了研究。但未见树体不同部位的光环境在核桃果实发育过程中对木质素与相关酶活性变化的影响方面的报道,本研究从树冠不同方位、不同冠层下核桃果实内果皮木质素积累及相关酶活性的变化规律进行探索分析,寻找影响核桃果实内果皮发育的因素,以期为后期核桃栽培管理中树形修剪和株行距的确定提供参考。

1 材料和方法

1.1 试验地概况

试验于阿克苏地区温宿县木本粮油林场新疆林科院良种核桃示范园内进行,温宿县位于天山中

段南坡一塔里木盆地北部边缘,地理坐标位于东经79°28'~81°30'、北纬40°52'~42°15'。当地的气候特点:降水量稀少,蒸发量大,昼夜温差悬殊,属典型的大陆性气候。年平均气温10.10℃,极端最低气温-27.4℃,太阳辐射年均总量140 kcal·cm⁻²,年平均日照时数为2 747.7 h,年均降水量65.4 mm,年均蒸发量956.3 mm,年均无霜期185 d。

1.2 试验材料

试验材料为新疆主栽核桃品种‘新新2号’,栽植株行距为3 m×6 m,东西行向,树龄为15 a(年),树高7~8 m,冠幅5~6 m,无病虫害,树势较开张。选取树势健壮,生长势相对一致的‘新新2号’核桃样株6棵,在每棵样树的同一部位采果3个后混合,取混合样分成3份作为3次重复。

取样时将核桃样树进行不同部位的划分,垂直分布上分别采摘样树下层(距树干基部2.5 m以下)、中层(距树干基部2.51~5 m)、上层(距树干基部5 m以上)的果实样品;水平分布上分别采摘样树中部(距中心树干1.5 m以内)、南面、北面(距中心树干1.5 m以上)的果实样品;于2019年6月5日至2019年7月26日,间隔7 d采样一次,共7次,每次采摘后根据不同部位将果实进行分类装袋标记,带回实验室-40℃低温冰箱保存,用于各项指标的测定。

1.3 试验方法

木质素沉积观察采用间苯三酚法^[16]。

木质素的测定采用硫酸法^[23](Klason法)。

苯丙氨酸解氨酶(PAL)的测定:参照陶书田等^[24]的方法。以每g鲜样光密度每h变化0.01为1个酶活性单位。

过氧化物酶(POD)的测定:参照陶书田等^[24]的方法。以每g鲜样光密度每min变化0.01为1个酶活性单位。

多酚氧化酶(PPO)活力的测定:参照吴有根等^[25]的方法。以每g鲜样光密度每min变化0.01为1个酶活性单位。

1.4 数据分析

采用Excel、SPSS19.0、Origin软件进行数据处理、制图。

2 结果与分析

2.1 核桃内果皮硬核发育期木质素沉积的变化

如图1所示,6月5日在内果皮顶端、内隔膜处已

有一部分被染成樱桃红色,表明内果皮木质化始于果顶端及内隔膜。此后木质素不断沉积,至6月23日内果皮中出现大部分着色,且颜色已转变为紫红色但相对较薄,表明此时核桃内果皮初步成型,坚果大小定

型。至7月9日,随着种仁逐渐形成内果皮着色层亦逐渐增厚,颜色也逐渐加深。表明硬壳厚度增加,木质素加速积累。至7月26日内果皮厚度不再增加,表明内果皮分化已经结束并已沉积大量木质素。



从左至右分别为核桃的纵切、横切、缝合线纵切,粉红色为木质素沉淀部位。

Walnuts from left to right in the picture are longitudinal section, cross section and longitudinal section of suture line. Pink section is the part of lignin deposition.

图1 硬核发育期木质素沉积的变化

Fig. 1 Changes in lignification of walnut endocarp during hardcore development

2.2 核桃内果皮中木质素含量的变化

对树冠不同方位、不同冠层果实的内果皮进行木质素含量测定,如图2所示,在果实发育过程中树

冠不同方位、不同冠层内果皮木质素含量呈现持续上升的趋势,说明随着核桃果实的发育,内果皮层不断分化,木质素沉积不断增加。冠层水平分布上,南

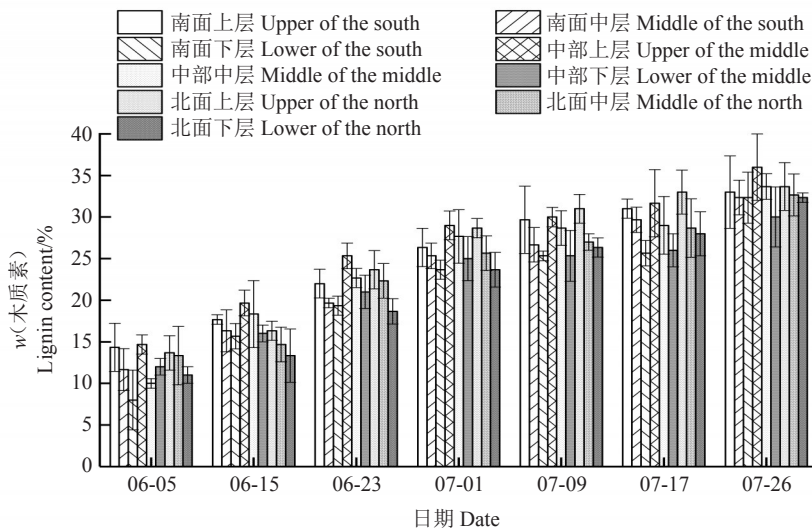


图2 不同部位核桃内果皮木质素含量的变化

Fig. 2 Changes of lignin in walnut endocarp in different positions

面、中部和北面三个方位的木质素含量差异不大,木质素含量高低依次表现为:南面>北面>中部,其中南面木质素含量稍高于北面,北面稍高于中部。在冠层垂直分布上,上层木质素含量高于中层和下层,木质素含量高低依次表现为:上层>中层>下层。在不同方位、不同冠层上,木质素含量高低的排序依次为:南面上层>南面中层>南面下层>北面上层>北面中层>北面下层>中部上层>中部中层>中部下层。

2.3 核桃内果皮中 PAL 酶活性的变化

对树冠不同方位、不同冠层果实的内果皮进行 PAL 酶活性测定,如图3所示,PAL 酶活性基本呈现先上升后缓慢下降的趋势。由6月5日的均值 $49.03 \text{ U} \cdot \text{g}^{-1}$ 上升到6月15日的均值 $118.96 \text{ U} \cdot \text{g}^{-1}$,之后 PAL 酶活性逐渐下降至均值 $52.87 \text{ U} \cdot \text{g}^{-1}$ 。不同方位、不同冠层对核桃内果皮 PAL 酶活性的影响不同,在冠层水平分布上,每个采样日期的南面酶活性均大于北面和中部,PAL 酶活性表现为:南面>北

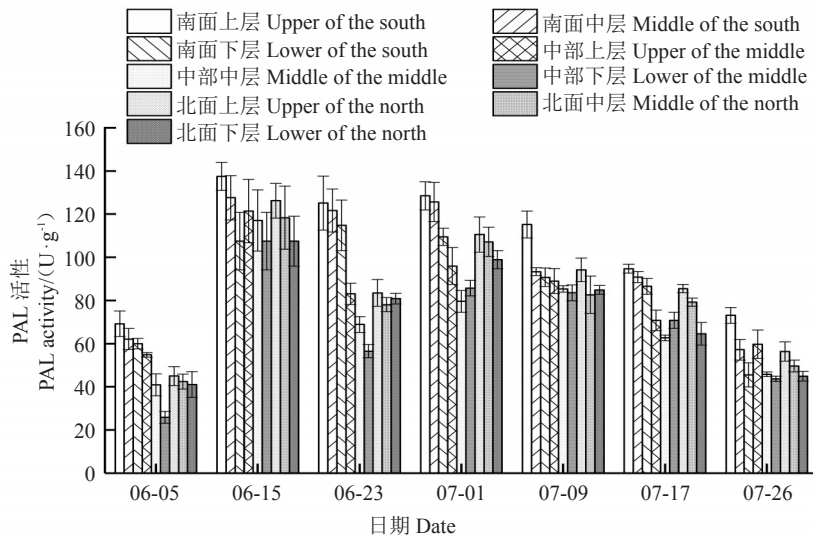


图3 不同部位核桃内果皮 PAL 酶活性的变化

Fig. 3 Changes of PAL activity in endocarp of walnut in different positions

面>中部。在冠层垂直分布上,每个采样日期的上层酶活性均大于中层和下层,PAL 酶活性表现为:上层>中层>下层;在不同方位、不同冠层上,PAL 酶活性强弱的排序依次为:南面上层>南面中层>南面下层>北面上层>北面中层>北面下层>中部上层>中部中层>中部下层。

2.4 核桃内果皮中 POD 酶活性的变化

对树冠不同方位、不同冠层果实的内果皮进行 POD 酶活性测定,如图4所示,POD 酶活性总体呈现先下降后上升再逐渐下降的趋势。两个高峰点出现在6月5日和6月23日,均值分别为: $39.99 \text{ U} \cdot \text{g}^{-1}$ 、 $33.53 \text{ U} \cdot \text{g}^{-1}$ 。不同方位、不同冠层对核桃内果皮 POD 酶活性的影响不同,在冠层水平分布上,每个采样日期的南面酶活性均大于北面和中部,POD 酶活性表现为:南面>北面>中部;在冠层垂直分布上,每个采样日期的上层酶活性均大于中层和下层,POD 酶活性表现为:上层>中层>下层。在不

同方位、不同冠层上,POD 酶活性强弱的排序依次为:南面上层>南面中层>南面下层>北面上层>北面中层>北面下层>中部上层>中部中层>中部下层。

2.5 核桃内果皮中 PPO 酶活性的变化

对树冠不同方位、不同冠层果实的内果皮进行 PPO 酶活性测定,如图5所示,PPO 酶活性呈现先下降后上升再逐渐下降后上升的整体趋势。不同方位、不同冠层对核桃内果皮 PPO 酶活性的影响不同,在冠层水平分布上,每个采样日期的北面酶活性均大于中部和南面,PPO 酶活性表现为:北面>中部>南面;在冠层垂直分布上,每个采样日期的下层酶活性均大于中层和上层,PPO 酶活性表现为:下层>中层>上层;在不同方位、不同冠层上,PPO 酶活性强弱的排序依次为:北面下层>北面中层>北面上层>中部下层>中部中层>中部上层>南面下层>南面中层>南面上层。

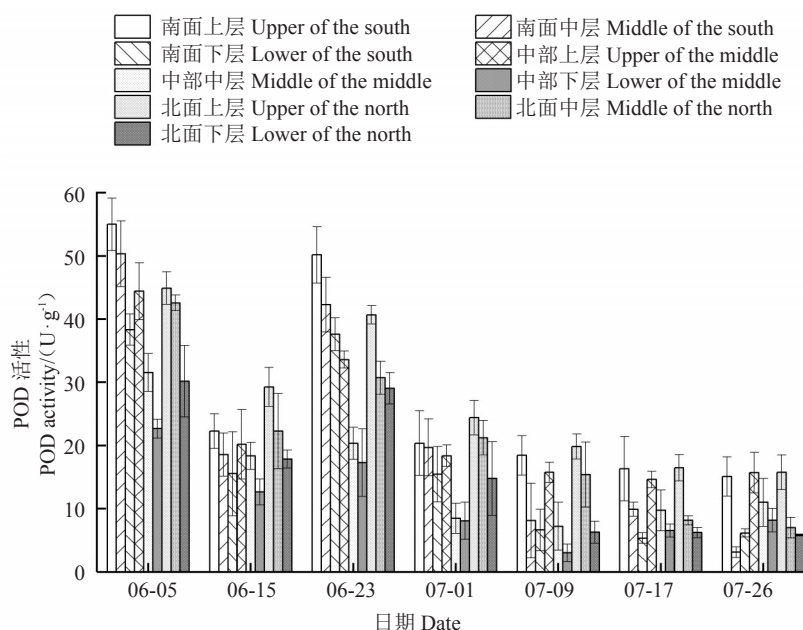


图4 不同部位核桃内果皮POD酶活性的变化

Fig. 4 Changes of POD activity in endocarp of walnut in different positions

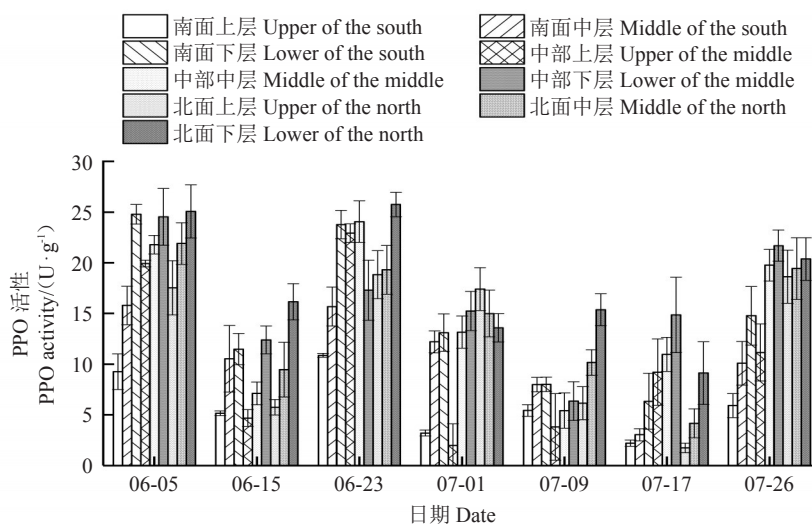


图5 不同部位核桃内果皮PPO酶活性的变化

Fig. 5 Changes of PPO activity in endocarp of walnut in different positions

2.6 核桃果实内果皮中木质素含量与相关酶活性变化的相关性分析

对核桃果实发育期的内果皮中PAL、POD、PPO酶活性与木质素进行相关性分析,如表1所示,木质素与POD酶活性呈极显著正相关($p < 0.01, r = 0.808^{**}$),POD酶活性与PAL酶活性呈显著正相关,与PPO酶活性极显著负相关($p < 0.01, r = -0.936^{**}$),PAL酶活性与PPO酶活性呈极显著负相关($p < 0.01, r = -0.842^{**}$)。

3 讨论

核桃硬壳发育即为内果皮薄壁细胞向石细胞分化过程,是细胞次生壁形成及增厚并不断木质化的过程^[26]。本研究中通过染色实验发现,木质素呈现出从内果皮两顶端开始向中间不断积累,颜色从樱桃红到紫红色的变化过程,且内果皮厚度和硬度不断增加。相关研究发现^[26-27],核桃果皮发育初期,内果皮细胞小而透明,内、中、外三层果皮的界线未

表1 核桃果实内果皮中木质素含量与相关酶活性变化的相关性分析

Table 1 Correlation analysis of lignin content and related enzyme activities in walnut fruit endocarp

	PAL	POD	PPO	木质素 Lignin
PAL	1			
POD	0.711*	1		
PPO	-0.842**	-0.936**	1	
木质素				1
Lignin	0.251	0.808**	-0.627	1

注: * 和 ** 分别表示在 0.05 和 0.01 水平(双侧)上显著相关。

Note: * and ** denote significance at 0.05 and 0.01 probability level (bilateral).

能区分, 后期内果皮细胞壁增厚, 逐渐转化为坚硬的木质化石细胞层。本研究中发现内果皮细胞层数随着果实体积的增大而不断增加, 内果皮颜色由质嫩的白色变成乳白色, 硬度也随之增强, 且内果皮外侧刻纹不断凸显; 后期随着木质素逐渐沉积, 着色层不断增厚。通过木质素沉积观察和木质素含量的测定, 发现木质素沉积由少到多、由薄到厚, 颜色由浅到深, 同时内果皮机械强度增加, 这与木质素含量随着核桃果实的发育呈持续上升的趋势同步。此结果与文菁等^[13, 17]的研究结果一致。

研究表明, 核桃内果皮的主要成分为木质素和纤维素^[28]。木质素是包围在木纤维等管束细胞和厚壁细胞壁外的一类物质, 其生物合成是通过苯丙烷类代谢途径进行^[29-31], 苯丙烷类代谢过程中有许多的酶参与, 其中 PAL、POD、PPO 在木质素形成过程中起着重要作用^[32-35]。PAL 是苯丙烷代谢途径中的关键酶和光诱导酶^[36]; POD 是催化木质素单体合成木质素的重要酶类, 影响组织的木质化^[37]; PPO 能够在酚类物质氧化过程中发挥作用, 有助于木质素前体物质的形成^[38]。本研究中 PAL、POD 酶活性在后期逐渐降低, PPO 酶活性反而出现升高的趋势, 据此可认为酚类物质在木质素后期的沉积中起着主要作用。这与李夕勃^[13]、文菁等^[17]、朱秋萍等^[34]的研究结果一致。由于核桃果实随着树冠层次由低到高的增加, 其受到的光照强度逐渐增强, 且由北向南受到的光照强度也依次增强^[39]。因此在冠层垂直分布上, PAL、POD 酶活性表现为上层 > 中层 > 下层, 相反 PPO 酶活性表现为下层 > 中层 > 上层; 这是由于在试验园内随着树冠层次的升高, 光照强度的增加, PAL、POD 酶活性增强; 而 PPO 酶活性随着光照

强度的增加而受到抑制; 表明了光照强度对核桃果实内果皮中木质素的积累及相关酶活性的增强起着诱导作用。这与刘小阳等^[40]、卢绍浩等^[41]、李丽琼等^[42]的研究结果相符。在冠层水平分布上, PAL、POD 酶活性表现为南面 > 北面 > 中部, 而 PPO 酶活性表现为北面 > 中部 > 南面; 这是由于在试验园内树冠不同方位所接受的光照强度存在差异, 即树冠南面接受的光照强度最强, 其次是中部、北面; 北面光照相对中部更弱, 但 PAL、POD 酶活性表现比中部更强, 这可能是由于树冠北面通风、空气湿度等微气候条件比树冠中部好, 从而形成北面酶活性高于中部, 具体原因有待下一步深入研究。

相关性分析表明, 木质素含量与 POD 酶活性变化呈极显著正相关, 与 PPO 酶活性变化呈负相关。这与前人^[17, 34]的研究结果不同之处在于, 前人是在不同品种间的硬壳主要成分与相关酶活性变化相关性分析, 本研究是在同一树体的不同方位、不同冠层中木质素含量与相关酶活性变化的相关性分析。因此, 相关性分析结果存在差异。

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