

植物生长调节物质对葡萄着色影响的研究进展

李芳菲^{1,2}, 马文瑶^{1,2}, 程大伟¹, 黄海娜¹, 顾红¹, 陈锦永^{1*}, 杨英军^{2*}

(¹中国农业科学院郑州果树研究所, 郑州 450009; ²河南科技大学, 河南洛阳 471023)

摘要:色泽是葡萄果实重要的品质特征,果实着色受诸多因素的影响,除基因型、气候条件、土肥水平和栽培技术外,植物生长调节物质也可影响果实着色。笔者对6种植物生长调节物质(脱落酸、萘乙酸、乙烯利、茉莉酸类、赤霉素、油菜素内酯)对葡萄着色的影响进行综述,除萘乙酸抑制葡萄果实着色外,其他物质均能促进果实着色和提前成熟。笔者从生理和分子水平阐述主要植物生长调节物质影响葡萄果实着色的作用机理,为外源生长调节物质在促进/抑制葡萄着色的应用方面提供理论依据。

关键词:葡萄;植物生长调节物质;着色;花色苷

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Advances in grape coloration regulated by plant growth regulators

LI Fangfei^{1,2}, MA Wenyao^{1,2}, CHENG Dawei¹, HUANG Haina¹, GU Hong¹, CHEN Jinyong^{1*}, YANG Yingjun^{2*}

(¹Zhengzhou Fruit Research Institute, CAAS, Zhengzhou 450009, Henan, China; ²Henan University of Science and Technology, Luoyang 471023, Henan, China)

Abstract: Fruit color is an important indicator of the quality of grape berry, which directly affects the nutritional and commodity value of grapes. Fruit coloration is determined by pigments, such as chlorophyll, carotenoids and anthocyanidins. The main pigment in the colored grape is anthocyanidin, which is present in the form of anthocyanins in grapes. Synthesis of anthocyanins is regulated by internal factors such as genotype, enzymes, hormones, and external factors such as light, temperature, water, soil nutrient and so on resulting in poorly colored fruit, thus affecting quality and commodity value. Therefore, it is necessary to take measures to promote coloration of grape. Using of plant growth regulators is undoubtedly a convenient and quick means. In this paper, the effects and mechanisms of different plant growth regulating substances on grape coloration were expounded from the physiological and molecular levels, including abscisic acid (ABA), naphthalene acetic acid (NAA), ethephon (ETH), jasmonates (JAs), gibberellic acid (GA) and brassinolides (BRs). On physiological level: 1. Promoting coloring by increasing sugar content. Coloration of the fruit depends on anthocyanin content. Sugar is an important base material for anthocyanins synthesis. ABA, JAs, BR, etc. can increase the accumulation of sugar in fruits, thereby promoting the formation of anthocyanins. However, NAA reduces the sugar content of the fruit, which is not conducive to anthocyanin synthesis. 2. Promoting coloring by affecting the pigment content of fruits. The use of exogenous plant growth regulators (ABA, ETH, JAs, GA, BRs) can induce chlorophyll degradation, promote the synthesis of carotenoids and anthocyanins, accelerate the color change of the peel and the maturation process, but NAA has the opposite effect. 3. Promoting coloration by regulating endogenous hormones. Plant growth regulating substances can regulate the con-

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作者简介:李芳菲,女,在读硕士研究生,研究方向为果实着色生理与技术。Tel:18848966598, E-mail:1193297033@qq.com

*通信作者 Author for correspondence. E-mail:chenjinyong@caas.cn; E-mail:Yangyingjun2003@126.com

tent of endogenous hormones and the balance between them, moreover, regulates fruit maturation by interaction of hormones. For example, exogenous ABA increases the content of endogenous ABA and ethylene, while reduces the content of indoleacetic acid (IAA), GA, and Zealin-riboside (ZR); Exogenous ETH treatment can increase endogenous ETH content in fruit and then promoting coloring; BRs can increase the content of endogenous ABA, promote the release of ETH, reduce the content of IAA in the early stage of grape ripening, so that they can promote the coloration and maturation of grapes. On molecular level: The biosynthesis of anthocyanins is controlled by the expression level of structural genes and transcription factors. Structural genes directly encode biosynthesis enzymes in anthocyanin biosynthesis pathway, including: chalcone synthases (CHS), chalcone isomerase (CHI), flavonoid 3-hydroxylase (F3H), flavonoid 3'-hydroxylase (F3'H), flavonoid 3',5'-hydroxylase (F3'5'H), dihydroflavonol 4-reductase (DFR), leucoanthocyanidin dioxygenase (LDOX) and UDP glucose-flavonoid 3-O-glucosyltransferase (UGT). Transcription factors are not directly involved in the formation of anthocyanins, however they can regulate the expression and pattern of anthocyanin biosynthesis genes while controlling the changes in anthocyanins in time and space. At present, three kinds of anthocyanin-synthesized transcription factors have been isolated and identified: R2R3-MYB protein, bHLH protein and WD40 protein. Plant growth regulators (ABA, ETH, JAs, BRs, etc) can regulate the coloration of grape fruit through increasing the expression of anthocyanin related structural gene and transcription factors. NAA down-regulates the expression of structural genes, inhibits the synthesis of anthocyanins and delays fruit ripening. Currently plant growth regulating substances are widely used in grape coloring, but there are still many problems such as the abuse of plant growth regulators by people eager to promote coloring using excessive concentrations, resulting in poor coloration, fruit detachment, fruit cracking, leaf browning and other side effects. In view of the above problems in practical application, the variety characteristics of grape, the scientific cultivation management measures and the mechanism of plant growth regulation substances should be combined to select suitable application concentration, period and method for achieving a comprehensive quality of fruit control and improvement.

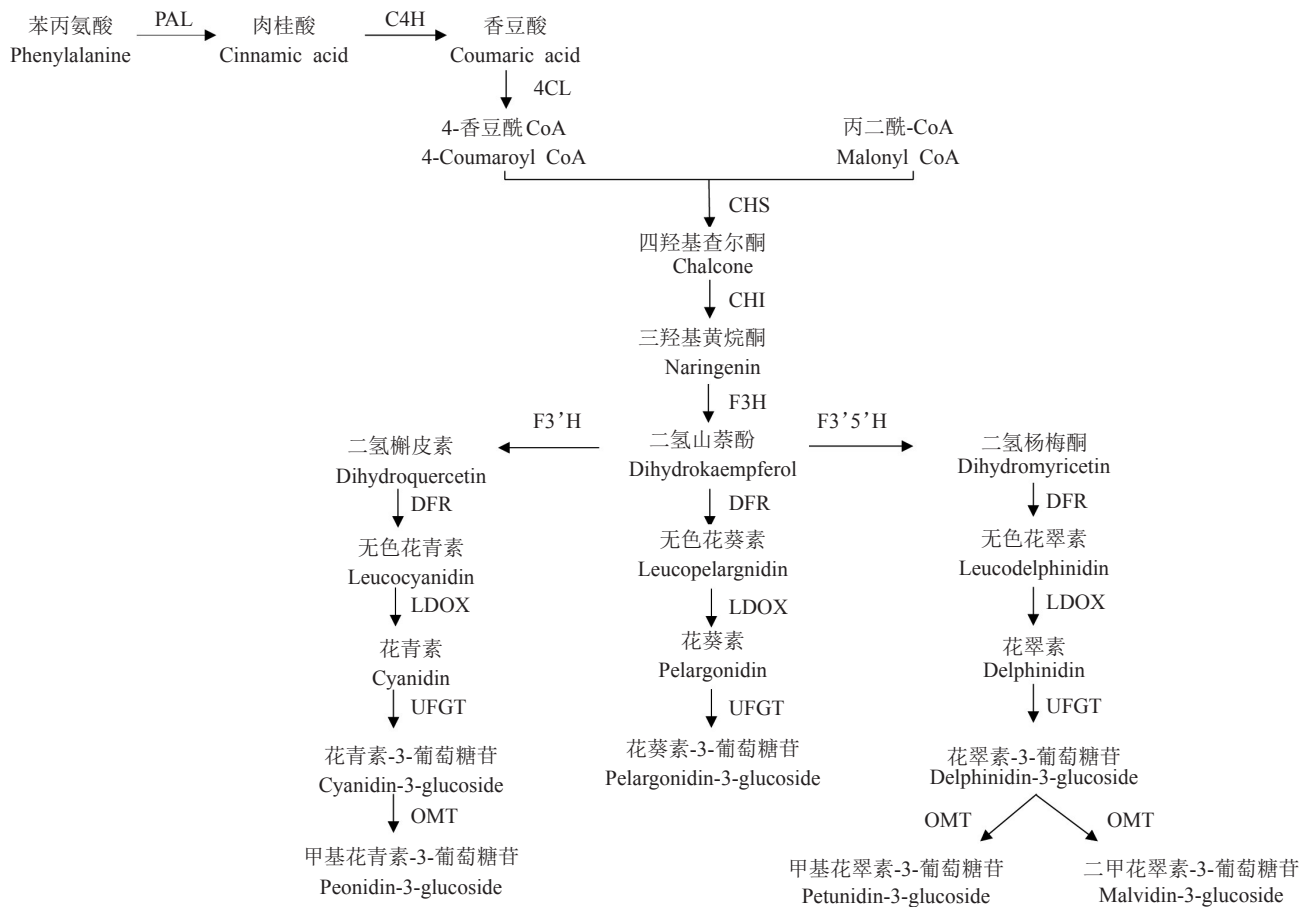
Key words: Grape; Plant growth regulating substances; Coloring; Anthocyanins

葡萄果皮色泽是衡量其商品价值的重要指标之一,直接影响葡萄的营养价值和加工性能^[1]。葡萄果皮中的色素以花色素类为主,包括花青素(cyanidin)、花翠素(delphinidin)、甲基花青素(peonidin)、甲基花翠素(petunidin)和二甲花翠素(malvidin)这五大类,通常以花色苷(糖苷)的形式存在。植物通过苯丙烷类路径和类黄酮路径合成花色苷,花色苷生物合成途径参考Azuma等^[2]并作修改(图1)。苯丙氨酸作为花色素合成的底物,经过苯丙烷类路径生成活化的4-香豆酰CoA,再经类黄酮途径形成无色花色素,然后经酶的催化最终形成花色苷,储存于液泡中^[3]。参与花色苷生物合成的基因分为结构基因和转录因子,结构基因可直接编码花色苷生物合成路径中的酶,如PAL、CHS、CHI、F3H、DFR、LDOX和UGT;转录因子不直接参与花色苷的合成,但它们可调控花色苷生物合成结构基因的表达

量和表达模式。在类黄酮的上游代谢途径中,不同家族的转录因子共同调节花色苷的形成,这些基因编码的蛋白通过结合相关结构基因的启动子从而控制结构基因的转录^[4]。目前已经分离和鉴定了3类花色苷合成的转录因子:R2R3-MYB蛋白;MYC家族的bHLH蛋白;WD40蛋白^[5]。

葡萄为非呼吸跃变型果实,其生长曲线表现为双S形,即在两个生长期(stage I和stage III)中间有一个缓慢期(lag phase, stage II)^[6]。由缓慢期过渡到生长III期,在葡萄栽培上称为转色期(veraison),也就意味着浆果开始成熟,整个过程伴随着糖分和花色苷的积累以及叶绿素的降解。基因型、生态和栽培条件都会影响葡萄果实花色苷的含量和各种花色苷的相对比例^[7-14]。

有色葡萄果实着色状况是影响果品价格和市场竞争力的重要因素。在一些产区,由于光照、温度、



PAL. 苯丙氨酸解氨酶; C4H. 肉桂酸 4-羟化酶; 4CL. 4-香豆酰 CoA 连接酶; CHS. 查尔酮合酶; CHI. 查尔酮异构酶; F3'H. 黄烷酮 3-羟化酶; F3'5'H. 类黄酮 3',5'-羟化酶; F3'H. 类黄酮 3'-羟化酶; F3'5'H. 类黄酮 3',5'-羟化酶; DFR. 黄烷酮醇 4-还原酶; LDOX. 无色花色素双加氧酶; UFGT. UDP 葡萄糖-类黄酮 3-O-葡萄糖基转移酶; OMT. O-甲基转移酶。

PAL. Phenylalanine ammonia-lyase; C4H. Cinnamate-4-hydroxylase; 4CL. 4-coumarate: CoA ligase; CHS. Chalcone synthase; CHI. Chalcone isomerase; F3'H. Flavanone 3-hydroxylase; F3'5'H. Flavanone 3',5'-hydroxylase; F3'H. Flavanone 3'-hydroxylase; F3'5'H. Flavanone 3',5'-hydroxylase; DFR. Dihydroflavonol 4-reductase; LDOX. Leucoanthocyanidin dioxygenase; UFGT. UDP glucose-flavonoid 3-O-glucosyltransferase; OMT. O-methyltransferase.

图 1 花色苷生物合成途径

Fig. 1 Anthocyanin biosynthesis pathway

湿度等气象因子不适,葡萄果实存在着色不良现象,影响果实的外观质量和商品价值。关于如何改善葡萄果实着色和增加葡萄果皮中的色素含量,除了通过栽培技术改变气候、树体养分改善和采收时间调整等方面进行考虑外,使用植物生长调节物质也是改善葡萄果实着色的有效方法之一,且成本低、收效快、效益高、节省劳动力^[15]。笔者就脱落酸(Abscisic acid, ABA)、萘乙酸(Naphthalene acetic acid, NAA)、乙烯利(Ethephon, ETH)、茉莉酸类物质(jasmonates, JAs)、赤霉素(Gibberellic acid, GA)、油菜素内酯类化合物(Brassinolides, BRs)等植物生长调节物质对葡萄着色的影响研究进展进行综述,以期科学使用植物生长调节物质改善果实着色提供参考。

1 ABA对葡萄着色的影响

ABA是植物体内一种重要的内源激素,通过信号传导途径调控植物体内的一系列代谢过程^[16]。ABA不但在植物抵抗外界胁迫时起重要的作用,而且在果实着色、改善品质等方面都有积极作用^[17]。从20世纪80年代起,国际上开始进行ABA对葡萄成熟和色素形成的调控研究。目前,日本、美国等在改善葡萄着色方面已广泛推广应用ABA制剂^[18],国内研究也表明,ABA处理对葡萄着色及改善果实品质有促进作用^[19-22]。

1.1 ABA通过提高糖含量促进着色

果实着色状况取决于花色苷含量,而花色苷累

积与糖分有关^[23]。糖分不仅决定着果实甜度,而且还是花色苷合成的重要原料。研究发现,在果实成熟过程中,随着果实中ABA含量的增加,糖含量也在增加,果皮色泽逐渐加深^[24-25]。一些学者认为,经ABA处理可以使果实中有机酸作为呼吸基质更多地被氧化分解转化为糖,糖含量的增加促进了花色苷的形成及果实着色^[26]。胡春霞等^[23]研究发现,ABA能促进葡萄果皮中可溶性糖的积累和花色苷的形成,从而促进果实着色,且花色苷含量与可溶性糖含量呈显著正相关,进一步验证了ABA可通过提高糖含量促进葡萄着色。

1.2 ABA通过影响色素含量促进着色

果皮颜色的呈现是叶绿素、类胡萝卜素、花色苷等色素综合作用的结果,花色苷对果实红色和紫色起决定作用,类胡萝卜素使果实呈黄色、橙色或红色,而叶绿素对红色有一定干扰或屏蔽作用。谢周等^[27]使用不同浓度的ABA处理提高葡萄果实着色的研究表明,ABA加速了果实中叶绿素的降解和花色苷的形成,且叶绿素与花色苷含量呈显著负相关。以上研究表明ABA可通过影响其他色素代谢而促进果实着色。

1.3 ABA通过调节内源激素促进着色

外源ABA是通过调节内源激素含量及激素间的平衡来促进葡萄果实着色,主要是提高了内源ABA含量,降低了吲哚乙酸(IAA)、GA、玉米素核苷ZR含量,提高ABA与(IAA、GA、ZR)的比值^[28]。其中ABA、IAA含量及二者比值与花色苷合成间的关系更为密切,二者可能直接作用于花色苷合成相关基因^[29]。

果实成熟与花色素合成增加均与乙烯释放量的增加有密切的关系,外源ABA能提高果实中乙烯合成基因的转录水平,进而提高果实的乙烯释放量,乙烯可通过影响细胞膜透性、增加糖分运输和积累,为反应提供底物或直接调节生理生化过程从而促进花色苷的合成,进而促进果实着色^[30]。

1.4 ABA通过影响花色苷合成相关基因的表达促进果实着色

有研究表明,ABA处理能够促进葡萄果皮中花色苷合成的结构基因和转录因子的表达,从而促进葡萄果实着色^[31-33]。目前为止,已经从葡萄上分离出20多个与花色苷合成相关的基因,其中多数基因编码的蛋白为R2R3MYB蛋白,说明R2R3-MYB家族

的一些转录因子特异的调节花色苷的积累^[34]。MYB主要是通过调节UFGT基因的表达影响花色苷的积累^[35]。MYBA1和MYBA2两个转录因子均参与了葡萄花色苷合成的调控,通过进一步研究发现它们是作用于UFGT基因的启动子区而使其表达^[36]。使用ABA处理果实可促进花色苷合成路径MYBA1、MYBA2和UFGT的转录,并增加了花色苷合成途径中的一些中间产物的含量^[33]。

PAL和CHS分别是葡萄花色苷生物合成过程中苯丙烷类代谢路径和类黄酮路径的第一个关键酶,在果实着色期,PAL和CHS活性与花色苷的合成紧密相关^[37]。经外源ABA处理后,果皮PAL活性显著增加,PAL、CHS、CHI基因随着花色苷合成大量表达^[20,38],Jeong等^[33]也得到了同样的结论。花色苷生物合成相关酶DFR可催化黄烷酮醇还原成无色花色素,LDOX可催化无色花色素转变为有色花色素^[39-40]。脱落酸可以提高葡萄果皮中F3H、DFR、LDOX等花色苷合成相关基因的转录水平,从而促进花色苷的合成,提高果实中花色苷的含量^[31]。UFGT是葡萄花色苷合成的关键酶,它催化花色素与糖结合成糖苷,使不稳定的花色素转化为稳定的花色苷^[41]。很多研究指出,不同品种葡萄经ABA处理后,均可提高UFGT的表达量及UFGT酶活性,与此同时,花色苷含量也明显上升^[42-43]。综上,ABA促进果实着色的作用机理是ABA通过提高花色苷合成相关的转录因子及结构基因的转录水平,促进相关酶活性使花色苷合成量增加。

2 NAA对葡萄着色的影响

NAA作为一种人工合成的植物生长调节剂,与植物本身的生长素吲哚乙酸(IAA)的作用特点和生理功能较为相似,具有促进细胞分裂与膨大,提高坐果率等功能,可经叶片、树枝的嫩表皮及种子进入植株体内,在促进果树开花、防止落果、提高果实产量以及推迟果实成熟等方面已得到广泛应用^[33]。在葡萄上的应用主要是疏花疏果,防止采前落果,提高坐果,促进生根等^[15]。

关于NAA影响果实着色方面的研究相对较少,且多数为抑制花色苷的合成,不利于果实着色。周莉等以200 mg·L⁻¹ NAA处理‘京优’葡萄,结果表明,处理后的果皮花色苷种类少于对照,不含甲基花青素-3,5-双葡萄糖苷,且果皮花色苷总含量在整个

转色期至采收期均低于对照,说明NAA能抑制果实的着色,推迟果实成熟^[44],这与赵权等^[45]在山葡萄上的研究结果一致。雷鸣^[26]以同样浓度NAA处理‘红地球’葡萄,处理组与对照的色泽明亮度L、红绿色差指标a、色泽强度C、色度角H及花色苷含量差异不显著,说明NAA不能促进果实着色与成熟。

此外,一些学者关于NAA对果实着色进行了更深入的研究,经NAA处理后,葡萄果皮花色苷合成相关结构基因(*CHSs*、*CHIs*、*F3Hs*、*F3'5'H*、*DFR*、*LDOX*、*UFGT*等)相对表达量低于对照,表明外源NAA处理抑制其表达^[44]。

3 乙烯利对葡萄着色的影响

乙烯能加速果实成熟以及衰老脱落等生理过程,被称为“成熟激素”。在一定条件下,乙烯利不仅可释放出乙烯,还能诱导植株产生乙烯。在果实成熟过程中,乙烯利能活化磷酸酯酶以及其他与果实成熟相关的酶,可以加速果实成熟和脱落^[46]。近年来大量研究表明,用乙烯利处理葡萄,可促进叶绿素降解和花色苷等类黄酮化合物合成,提高果实着色^[47-51],但其促进着色与造成落叶落果的使用浓度较为接近,生产上适宜的用量还需进一步研究。

3.1 乙烯利通过影响果实色素含量促进着色

乙烯利能加速葡萄的呼吸速率,诱导叶绿素降解,促进类胡萝卜素和花色苷的合成,促进果皮颜色改变,加速成熟进程^[51]。国外学者研究发现,外源乙烯能提高果实内源乙烯含量及花青素、花翠素、甲基花青素、甲基花翠素、二甲花翠素等花色苷含量,加速果实颜色转变^[50]。相反,利用乙烯合成抑制剂或乙烯受体抑制剂可以延缓葡萄成熟进程,如1-甲基环丙烯(1-MCP)处理显著降低始熟期葡萄果实花色苷含量^[52-53]。

3.2 乙烯利通过影响内源乙烯含量促进着色

葡萄果实开始进入转色期前内源乙烯迅速合成,此过程启动了随后的果实迅速膨大、花色苷大量积累以及果实酸度下降^[52]。葡萄果实为非呼吸跃变型,果实内乙烯含量很少,外源乙烯处理能提高果实内源乙烯含量,促进着色。用乙烯利处理酿酒葡萄品种‘赤霞珠’(Cabernet Sauvignon)24 h,可使果实内源乙烯含量提高6倍^[54]。外源乙烯能够诱导乙烯合成相关基因1-氨基环丙烷-1-羧酸(ACC)合成酶基因*ACS*和ACC氧化酶基因*ACO*的表达、*ACO*酶

活性变化,增加内源乙烯生成量,诱导叶绿素的代谢^[55]。

3.3 乙烯利通过影响花色苷合成相关基因的表达促进果实着色

乙烯利可以促进花色苷合成相关基因的表达,从而提高葡萄果实着色^[48,50]。转录因子控制花色苷生物合成过程中相关结构基因的时空表达,影响花色苷生物合成的强度和模式^[56]。MYB类转录因子主要调节花色苷合成关键基因*UFGT*的表达,控制花色苷的积累,在葡萄果实着色过程中起重要作用^[57]。于淼等^[58]研究表明,在葡萄果实着色期,*MYBA1*、*MYBA2*和*UFGT*随花色苷的合成均上调表达,乙烯利促进花色苷合成的同时也提高*MYBA1*、*MYBA2*和*UFGT*的转录水平,且*MYBA1*的转录水平与花色苷合成呈显著正相关,与Jeong等^[33]研究结果相似。

对于结构基因*CHS*、*F3Hs*、*DFR*、*LDOX*和*UFGT*而言,大量研究表明用乙烯利处理可以在不同程度上提高它们的转录水平,进而促进花色苷合成。在葡萄着色期用乙烯利处理果穗,发现花色苷合成相关基因的转录水平都高于对照,尤其是*CHS*、*GST*、*UFGT*转录水平显著提高^[58];此外乙烯利还可以促进葡萄果皮中*DFR*、*LDOX*以及*F3Hs*的表达^[33]。*UFGT*是花色苷合成的关键酶,Kobayashi等^[59]发现,乙烯利处理可同时促进花色苷合成和*UFGT*的表达,进而促进果实着色。

4 茉莉酸类物质对葡萄着色的影响

茉莉酸类物质(JAs)是一种广泛存在于植物体内的新型植物生长调节物质,包括茉莉酸(JA)、茉莉酸甲酯(MeJA)、茉莉酸丙酯(PDJ)及其衍生物。JAs具有广谱的生理效应,在植物生长发育、抗逆性以及次级代谢产物的合成等方面都具有重要的调控作用^[60]。目前,茉莉酸酯类在葡萄上已得到应用^[61-62],发现其不仅能够提高作物体内总酚、类黄酮等次生代谢物的含量,还可以通过促进果实中次生代谢产物花色苷的积累,改善果实色泽,提高果实品质。

4.1 MeJA对葡萄着色的影响

MeJA作为茉莉酸的衍生物,具有较强的挥发性,且不易被离子化,能有效透过植物细胞膜,易在植物体内运输^[63]。因此,外源MeJA易进入植物体内

发挥信号传导作用,调节植物体一系列生理生化反应,进而促进果实着色,提升果实品质。

葡萄中的可溶性固形物90%以上是可发酵的糖类,糖可为花色苷的合成提供原料。研究发现,经MeJA处理的葡萄随着可溶性固形物含量的上升,果皮中花色苷不断积累,且可溶性固形物积累时期与花色苷积累时期相吻合,推测MeJA处理促进葡萄花色苷的生物合成可能通过促进果实中糖类积累,增强UFGT酶活性进而改善果实色泽^[60]。孙晓文^[60]用MeJA处理‘圣诞玫瑰’结果表明,MeJA能够显著降低叶绿素a、b含量,提高果皮中花色苷含量,进而促进着色。

花色苷合成是多种酶共同作用的结果。DFR、LDOX、UFGT作为花色苷合成路径下游催化酶,在红色葡萄果实着色过程中发挥重要作用。MeJA可以诱导葡萄花色苷合成基因DFR、LDOX、UFGT的表达,促进花色苷合成中DFR、LDOX、UFGT的活性,进而触发花色苷的有效积累,使花色苷累积增加^[64-66]。此外,MeJA可通过上调苯丙氨酸途径中PAL、ANS、MYB等基因的表达促进花色苷生物合成,进而对色泽产生影响^[67-68]。

4.2 PDJ对葡萄着色的影响

PDJ是一种人工合成的茉莉酸类衍生物。与MeJA相比,PDJ具有化学稳定性好、生理效应持续时间长和易被植物吸收的特点,更具有实用价值。在美国、日本等国家和地区的一些产区,PDJ已经应用到苹果和葡萄生产中,促进花色苷积累,改善果实色泽^[69]。

与MeJA作用相同,PDJ也能促进果实中的糖分积累,加速叶绿素降解,上调花色苷合成结构基因DFR、LDOX、UFGT以及转录因子MYB的表达,直接或间接促进花色苷的合成与积累,提高果实花色苷含量,促进果实着色^[60]。但在相同浓度下,PDJ对改善葡萄果实色泽、提高果实品质的作用效果优于MeJA,可能是由于PDJ作为合成的茉莉酸类物质(JAs),其C9-C10位的烯键被氢饱和,羧基被丙基酯化,化学稳定性好,比MeJA更易被植物吸收,且不易分解失效^[70],因此具有更好的应用效果与实用价值。

5 GA对葡萄着色的影响

GA是调节植物生长发育的五大激素之一,它

的主要生理功能是促进细胞伸长,诱导开花,打破休眠,促进雄花分化^[71]。目前,GA在葡萄生产中应用较为广泛,主要涉及葡萄无核化栽培、提高坐果率、促进无籽葡萄果实生长、增加果穗重、拉长葡萄穗轴长度,改善果穗紧密度等方面^[71-75]。

近年来,一些学者对GA促进葡萄着色进行了探究。赵荣华等^[76]以‘丽红宝’葡萄为试材,结果发现,花前7 d使用质量浓度为25 mg·L⁻¹的GA处理花穗,结合花后10 d使用质量浓度为100 mg·L⁻¹的GA处理果穗,能显著增加果皮中类胡萝卜素含量和花色苷含量,果实色泽提高。但花后使用GA质量浓度超过100 mg·L⁻¹时,会使果实中花色苷含量降低,造成果实着色不良,延迟成熟。白世践等^[77]研究表明,花后7 d使用质量浓度为20 mg·L⁻¹的GA处理‘克瑞森无核’葡萄果穗,其在果实外观及内在品质方面效果最佳,GA质量浓度超过20 mg·L⁻¹,其着色效果不好,分析是高浓度GA延迟了成熟。在开花前20 d及15 d使用3~7 mg·L⁻¹的GA处理‘西拉’葡萄花序,均可拉长果穗长度,降低坐果率及果穗质量,提高了葡萄花色苷含量,有利于果穗内部葡萄果实的着色,可能是由于GA使果穗疏松程度增加、通风透光条件改善、植株负载量降低,利于光合产物在生殖器官中积累^[78]。以上研究中GA促进葡萄着色的最适浓度相差较大,可能与品种的基因型有关。

关于GA对葡萄着色的研究表明,在一定浓度范围内使用GA处理葡萄果穗(花序),可提高果实品质,促进着色,而超过一定浓度范围则会延迟果实成熟,不利于着色,需要进一步探究其内在机制。

6 BRs对葡萄着色的影响

在作物中应用的油菜素内酯类化合物(BRs),主要有油菜素内酯(BR)和表油菜素内酯(EBR)^[79]。油菜素内酯类化合物(BRs)又被称作芸薹素、油菜素,是从植物花粉中提取出来的一种具有较强生理活性的新型植物生长调节物质^[80],广泛应用于果树产业。BRs在葡萄上的研究报道很多,应用效果表明BR能明显提高葡萄的坐果率和增加可溶性固形物含量,降低含酸量,促进葡萄着色与成熟,对葡萄的产量和品质都有一定提高^[81-83]。

6.1 BRs通过增加糖含量促进着色

葡萄果实BRs含量变化与ABA含量变化趋势相似,在葡萄幼果期,BRs含量较高,至果实转色前

其含量降到最低,当果实开始进入成熟期,BRs 含量迅速增加,并在果实成熟过程中一直保持在较高水平^[82]。外源 BRs 处理葡萄果实,可增加果实中可溶性糖的含量,促进花色苷积累,使果实提前转色和成熟^[23]。周秀梅等^[80]研究发现,BR 可以显著提高葡萄坐果率,且其单果粒重及果肉糖度等均有不同程度的提高。徐如涓等^[79]于花前一周和盛花期以 EBR 叶面喷施‘巨峰’和‘玫瑰香’葡萄,果皮全部转红率提高两倍以上,果实着色提早,可溶性固形物、总糖及游离氨基酸含量均比对照明显增加。以 BR 处理‘京亚’葡萄,其浆果色素含量显著提高,着色期提前,可溶性固形物含量提高,而含酸量下降,糖酸比明显增大^[81]。而使用 BRs 抑制剂 Brz 处理葡萄果实,则会降低果肉中可溶性糖的含量,延迟果实的成熟和着色^[82]。

6.2 BRs 通过影响内源激素促进着色

BRs 可提高内源 ABA 含量,促进乙烯释放,降低葡萄成熟初期果实的 IAA 含量,从而促进葡萄着色与成熟。外源 EBR 处理促进了葡萄果实花色苷的合成,同时提高了果实内源 ABA 含量;而 Brz 处理抑制了葡萄果实花色苷的合成,同时降低了果实内源 ABA 含量。Brz 处理‘赤霞珠’葡萄,其果实 ABA 含量高峰出现的时间也明显晚于对照和其他处理,说明果实 EBR 含量变化能影响 ABA 的合成与积累,但 BRs 是否能够直接作为信号分子来调控葡萄成熟,仍需进一步研究^[84]。BRs 在葡萄上的研究表明,BRs 能够提高乙烯合成过程中的关键酶 ACC 和 ACO 的活性,从而提高乙烯的释放量,促进果实着色^[85]。而 Brz 处理则降低了果实 ACO 活性和乙烯释放量,同时果实着色也被推迟^[84]。以上研究表明,在葡萄果实成熟过程中,BRs 可能与乙烯或 ABA 共同调控葡萄果实成熟。

6.3 BRs 通过影响花色苷合成相关基因的表达促进果实着色

BRs 可影响果实花色苷合成的调节因子和结构基因进而促进花色苷合成,促进葡萄着色。使用外源 EBR 处理可使‘赤霞珠’葡萄果皮花色苷合成的相关基因在转色期之前触发表达,类黄酮代谢途径中的调节因子 *MYBA1* 和 *MYBA2* 以及下游基因 *F3H*、*F3'H*、*F3'5'H*、*DFR*、*ANS* 和 *UFGT* 的表达与对照相比提前 14 d,上游基因 *CHS* 和 *CHI* 的表达提前 7 d。因此,EBR 处理对花色苷合成过程中两个调节

因子及下游基因的促进效果更显著^[21]。EBR 处理使‘赤霞珠’葡萄完全着色提前约 7 d,这个时间与处理对上游基因的促进时间一致,因此上游基因的表达对花色苷的合成非常重要,EBR 处理使‘赤霞珠’葡萄果皮完全着色提前的时间可能受上游基因(*CHS* 或 *CHI*)提前表达时间的限制^[86]。

7 问题和展望

果皮色泽是葡萄果实重要的感官指标之一,但在实际生产中会因为各种因素造成葡萄着色不良。植物生长调节物质已被广泛应用于葡萄产业,能够在一定程度上促进果实糖分积累,调控内源激素的水平和花色苷合成相关调节因子及结构基因的表达,进而影响葡萄果实的着色和成熟。

目前,植物生长调节物质在促进葡萄着色方面的研究与应用还存在诸多问题:使用植物生长调节物质时,忽略了栽培管理的重要性,影响了植物生长调节物质的使用功效;有些植物生长调节物质还存在毒性和残留问题;滥用植物生长调节物质,急于促进着色,或使用浓度过高,造成着色不良、果粒脱落、裂果以及叶片焦黄等副作用,例如乙烯利处理促进着色的浓度与其造成落叶落粒的浓度较为接近,在实际应用时要慎重。

针对以上问题,未来应加强新型植物生长调节物质的研制,以确保其安全性和应用效果。在实际应用中,应结合葡萄的品种特性、科学的栽培管理措施以及植物生长调节物质的作用机理,选择合理的施用浓度、施用时期和施用方法,以实现果实综合品质的调控和提高。

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